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# AI-BASED SOLUTION TO THE OPTIMIZATION PROBLEM OF THE MICROFLUIDIC GEOMETRY DESIGN

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2024, Novosibirsk, Russia

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# AI-BASED SOLUTION TO THE OPTIMIZATION PROBLEM OF THE MICROFLUIDIC GEOMETRY DESIGN

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# Report Outline

- 01** Review: MICROFLUIDICS FOR SINGLE RBC STUDIES (490 REFS)
- 02** CHIP DESIGN; EXPERIMENT; SINGLE CELL ANALYSIS TEST
- 03** TOP. OPT. 1: OF MINIMIZATION. MANUAL CREATION/TESTING OF NEW GEOMETRIES
- 04** TOP. OPT. 2: AUTO CREATION/TESTING OF NEW GEOMETRIES (10, 000 DATASETS).
- 05** TOP. OPT. 3: EA, AUTO CREATION AND IMPROVEMENT OF 300 NEW GEOMETRIES
- 06** TOP. OPT. 4: EA, AUTO CREATION SELF-IMPROVEMENT OF 30, 000 NEW GEOMETRIES
- 07** EXPERIMENT
- 08** CONCLUSION AND CLOSING REMARKS

# Roadmap Infographics



**Review:**  
**MICROFLUIDICS FOR**  
**SINGLE RBC STUDIES**  
**(492 REFS)**

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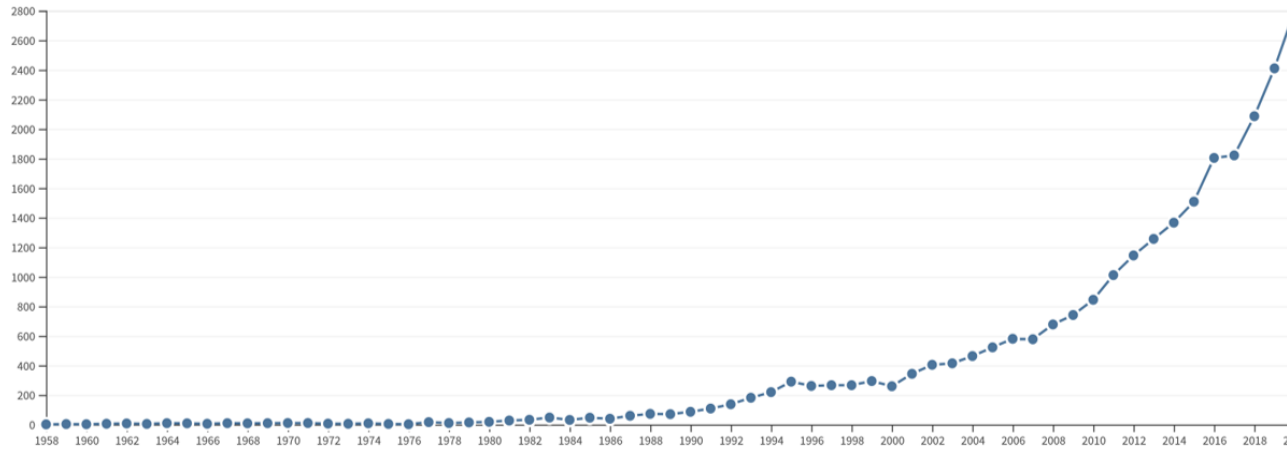
# RBC SCA: Attention rises



Public  
Single

RBC & single cell analysis  
967 publications (last 5 years)  
5,101 citations (last 5 years)

Sum of Times Cited per Year



90s 2000s 2010s





# Leading Causes of Death



Deaths for leading causes of death

59,041

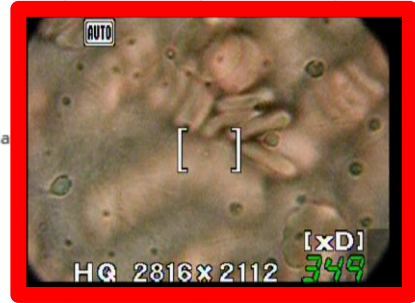
- Accidents (unintentional injuries): 173,040
- Chronic lower respiratory diseases: 156,979
- **Stroke (cerebrovascular diseases): 150,005**
- Alzheimer's disease: 121,499



otic syndrome, and nephrosis: 51,565  
 neumonia: 49,783  
 harm (suicide): 47,511

the United States, 2019, data table for figure 2

## Leading causes of death globally



Source: WHO Global Health Estimates.



# Problem Statement



## **uFD Chip.**

Biomicrofluidic chip, suitable for visual, quantitative and qualitative study of single RBC inside and outside.

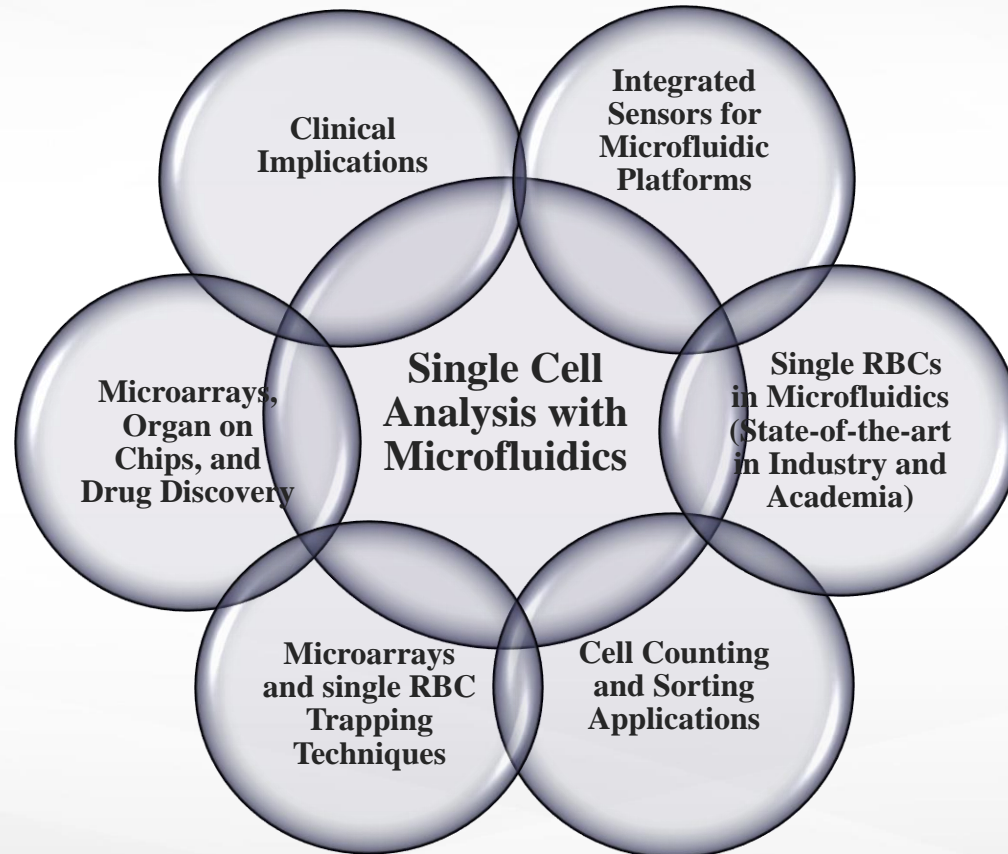
## **Trapping geometry.**

Trap RBCs in channels while keeping them suspended to allow fluidic flows around the cell..

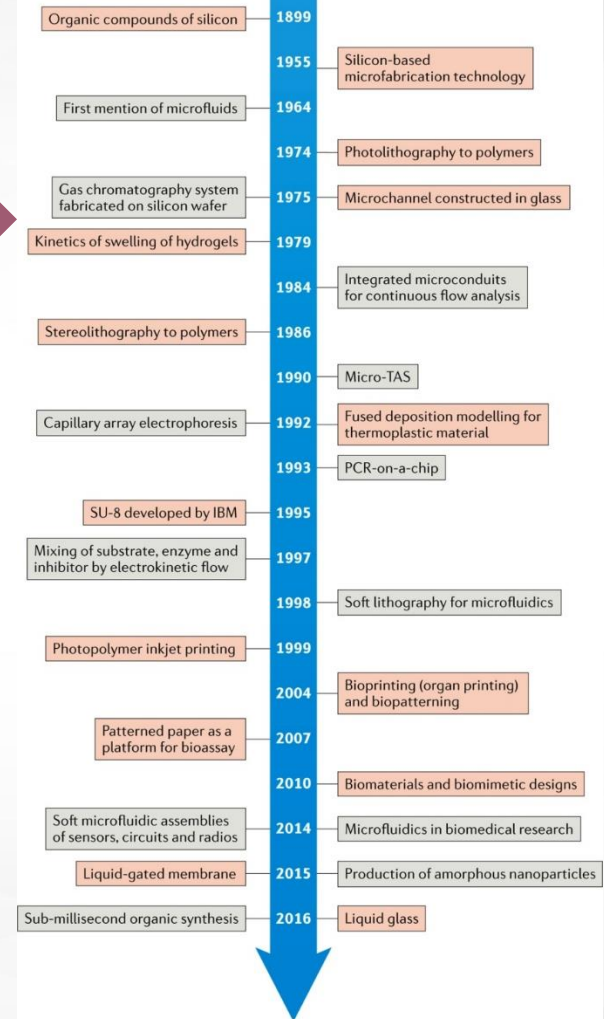
## **Topology Optimization.**

Achieve trapping efficiency 75%.





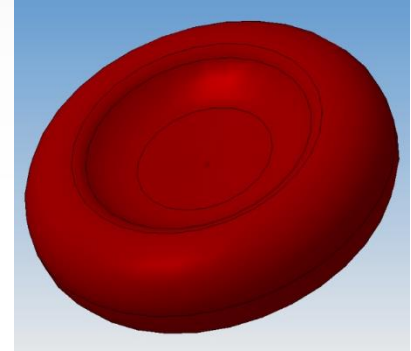
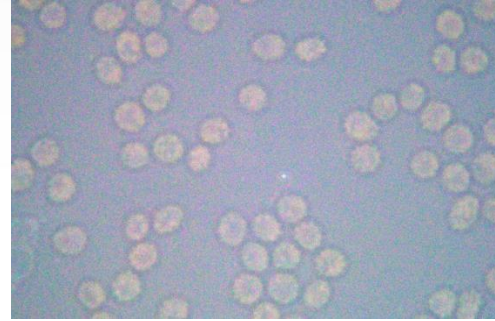
# Historical timeline of developments in materials and microfluidics



Hou, X. (2017) Interplay between materials and microfluidics  
*Nat. Rev. Mater.* doi:10.1038/natrevmats.2017.16

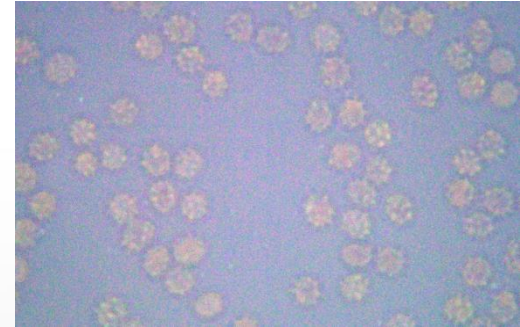
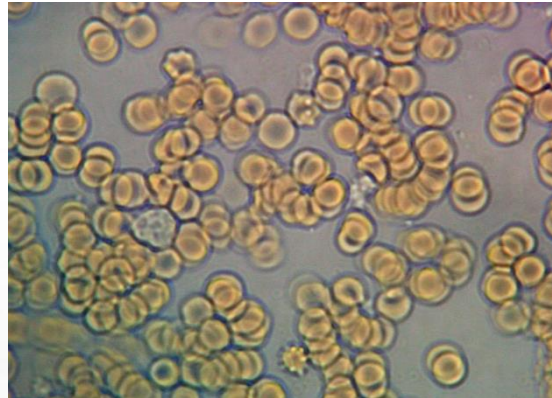


# Erythrocyte / RBC



A mature erythrocyte has a biconcave discoid shape. An erythrocyte is composed of hemoglobin (32%), water (65%), and membrane components (3%) and does not contain any nucleus.

Mazeron, S. Muller, and H. El. Azouzi, "Deformation of erythrocytes under shear: a small-angle light scattering study," *Biorheology* 34, 99-110 (1997).



$$z^2 = \left(\frac{0.86d}{2}\right)^2 \left[1 - \left(\frac{2x}{d}\right)^2\right] \left[0.01384083 + 0.2842917\left(\frac{2x}{d}\right)^2 + 0.01306932\left(\frac{2x}{d}\right)^4\right].$$



# RBCs differ:

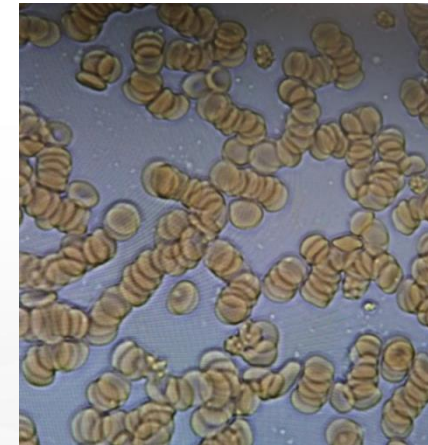
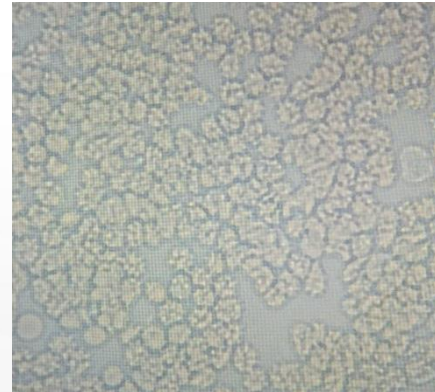
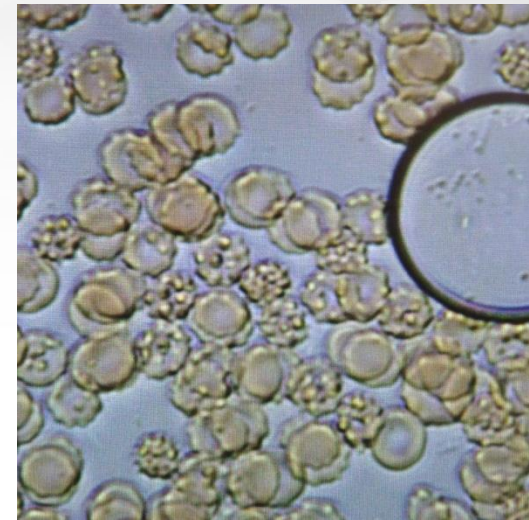
Depending on the value of the RBC's SC erythrocytes have different

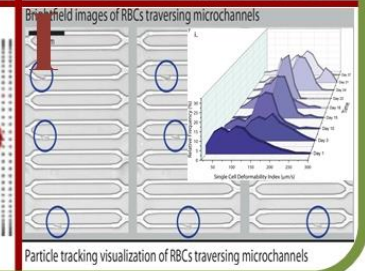
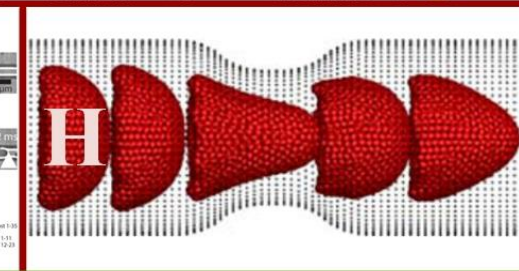
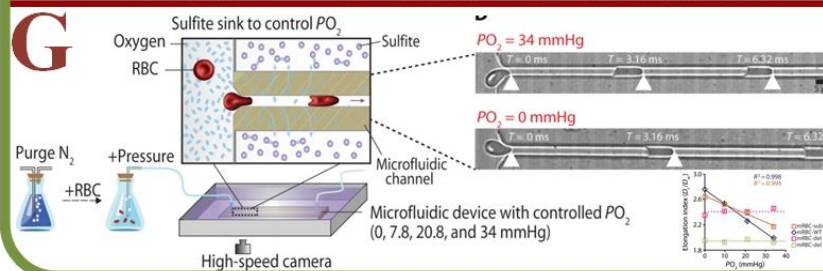
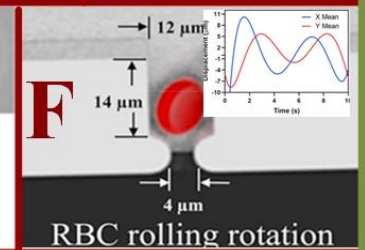
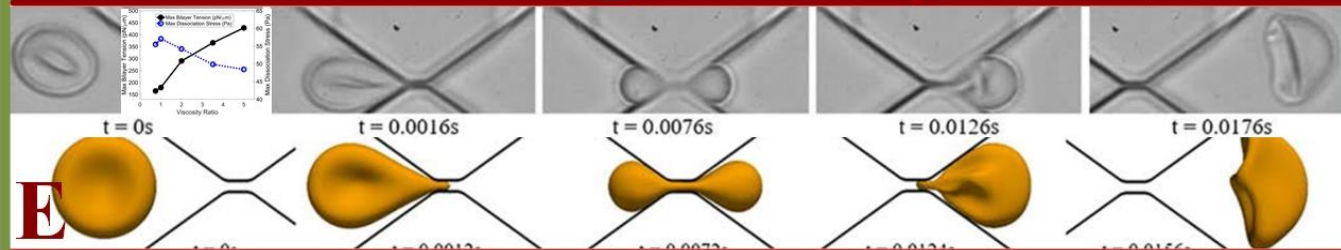
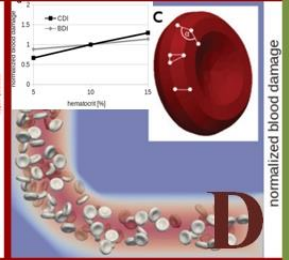
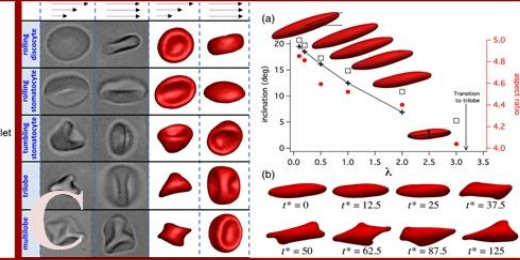
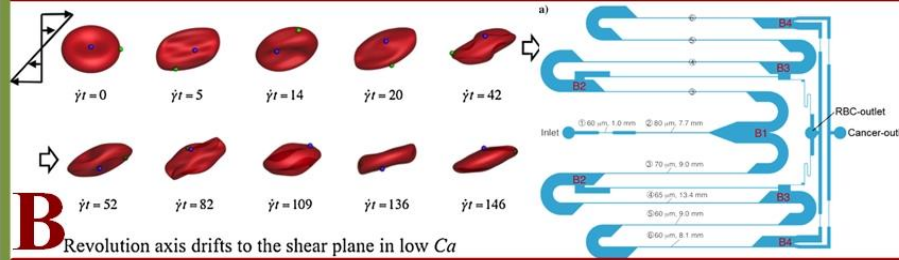
## shape:

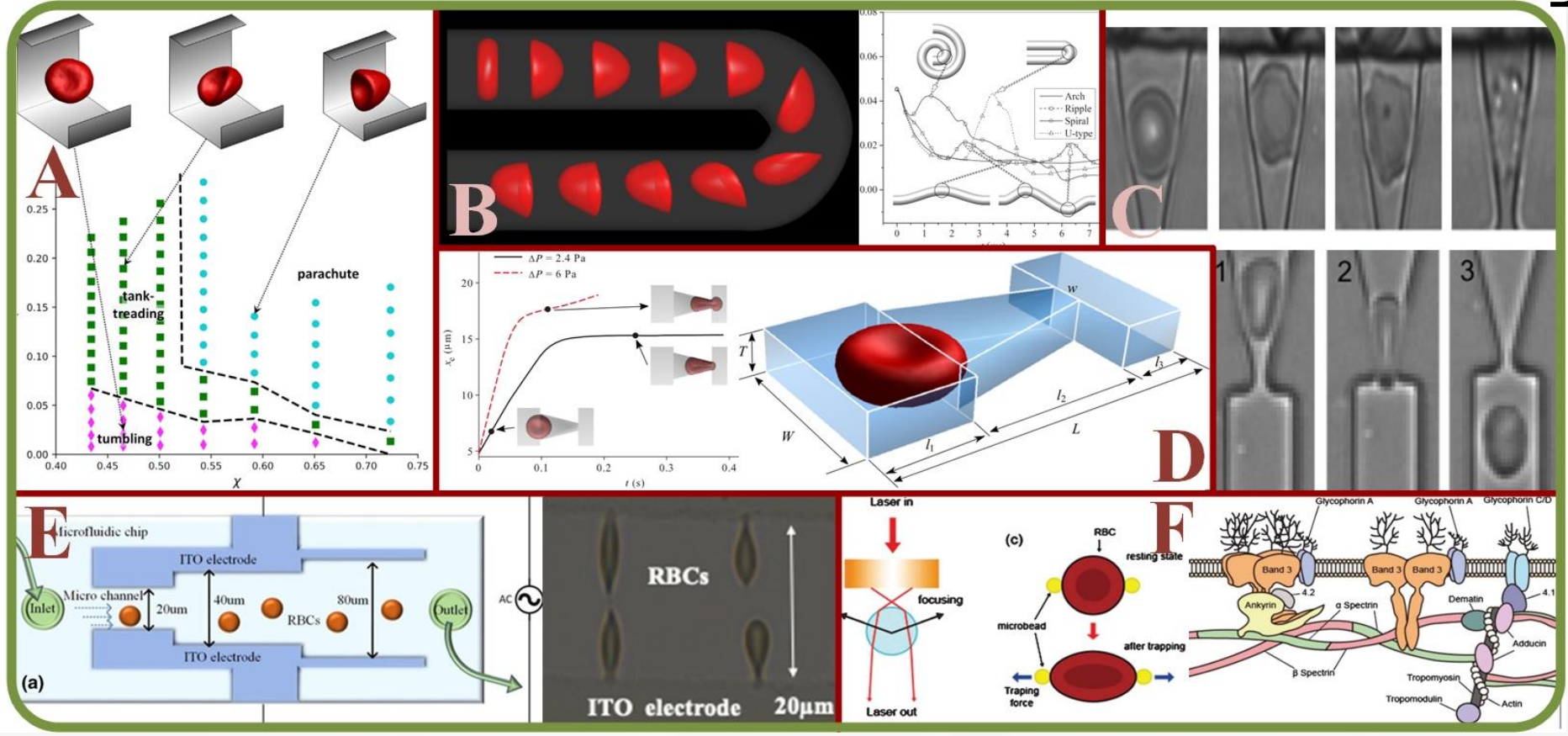
- Spherocytes
- Elliptocytes/Ovalocytes
- Stomatocytes
- Schistocytes
- Keratocytes
- Helmet Cells
- Acanthocytes
- Echinocytes
- Target Cells (Codocytes)
- Tear Drop Cells (Dacryocytes)
- Sickle Cells (Drepanocytes)
- Degmacytes

## size:

- Normocytes
- Anisocytosis
- Microcytes
- Macrocytes
- Megalocytes

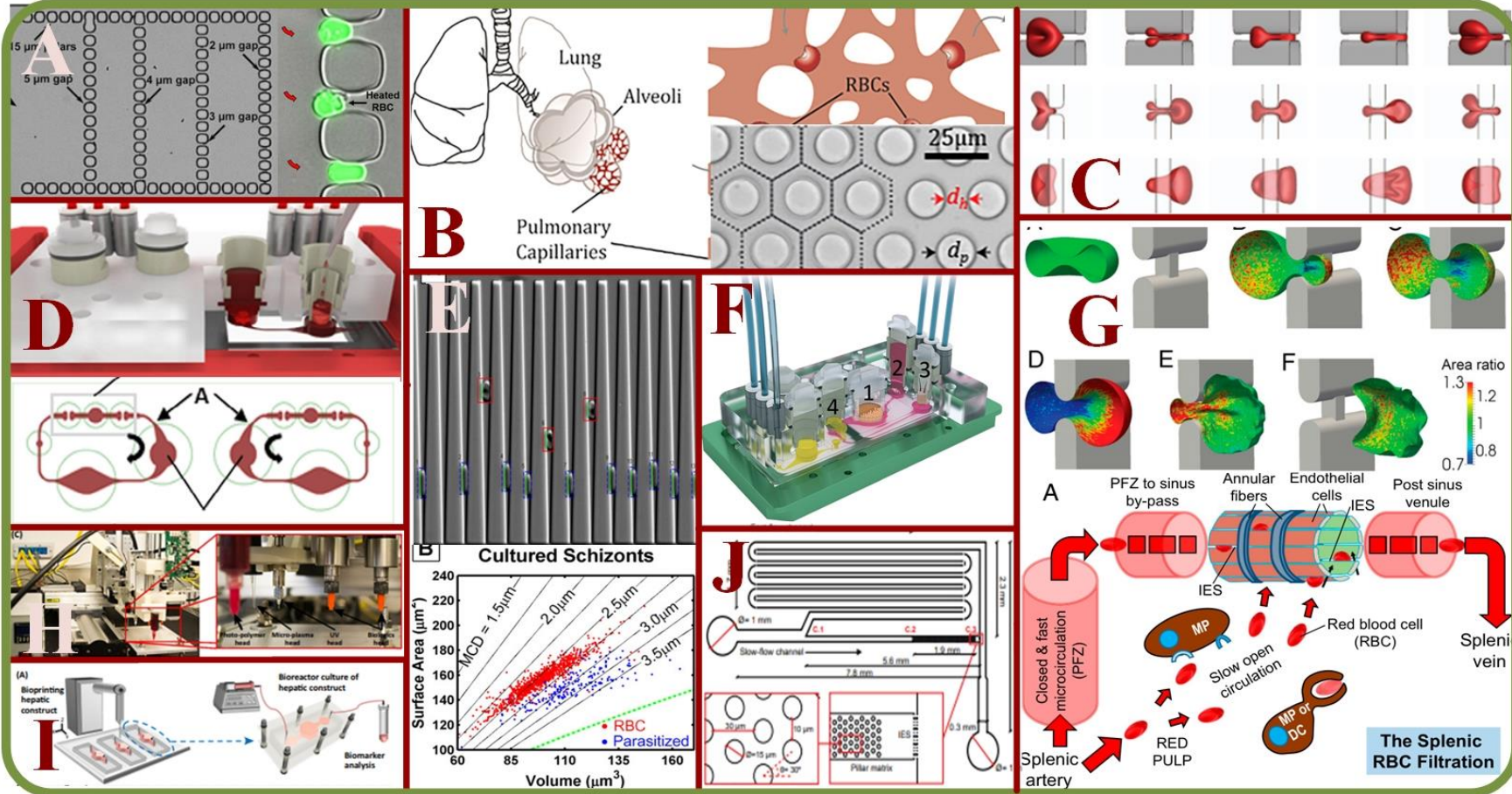






# Deformation of single Erythrocytes in Microchannels.





March 25, 2013

BOSTON – The Wyss Institute for Biologically Inspired Engineering at Harvard University announced today that it was awarded a **\$9.25 million** contract from the Defense Advanced Research Projects Agency (DARPA) to further advance **a blood-cleansing technology** developed at the Institute with prior DARPA support, and help accelerate its translation to humans as a new type of sepsis therapy.



# Funding

theranos

estimated value = 9,400,000,000\$  
USA Moon project = 20,400,000,000\$

the board of the directors members (all-star board):

- William **Perry** (former Secretary of Defense)
- Henry **Kissinger** (former Secretary of State)
- Sam **Nunn** (former U.S. Senator)
- Bill **Frist** (former U.S. Senator and heart-transplant surgeon)
- Gary **Roughead** (Admiral, USN, retired)
- James **Mattis** (General, USMC)
- George P. **Schultz** (former Secretary of State)
- Richard **Kovacevich** (former Wells Fargo Chairman and CEO)
- Riley **Bechtel** (chairman of the board and former CEO at Bechtel Group)
- William **Foege** (former director U.S. Centers for Disease Control and Prevention) and others



# Table 1 : Common Microfluidic Separation Methods

Method	Advantages	Disadvantages
Physical filtration	Increased cell separation and sorting efficiency, easily integrated with PDMS structures	Clogging and fouling of blood cells requires precise control of filter geometries
Hydrodynamic and hemodynamic processes	Inertial focusing for enhanced cell separation and sorting narrowed sheathed flows	Produce stress on cell samples, may alter molecular mechanisms, inhomogeneity
Surface Affinity and Topography	Specificity, cell purity	May alter cell physiology after sorting and isolation processes
Magnetophoresis	Directly differentiate blood cells without additives, efficiency up to ~ 90 %	Weak magnetic flux gradients on cells
Electrical Methods and Acoustophoresis	Sensitive, rapid, convenient, and robust. Easily integrated	Electrolysis, temperature elevations, phenotypic changes



# Innovation

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# Research Objectives

## Objective #1, uFD Chip



Biomicrofluidic chip, suitable for visual, quantitative and qualitative study of single RBC inside and outside

## Objective #2, Trapping geometry



Trap RBCs in channels while keeping them suspended to allow fluidic flows around the trapped cell.

## Objective #3, Topology Optimization



Achieve trapping efficiency 75%

CHIP DESIGN;  
EXPERIMENTAL TEST OF  
CAPABILITIES FOR  
SINGLE CELL ANALYSIS

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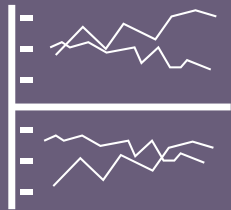


# Experimental results and discussion



## Local Optical Tomography

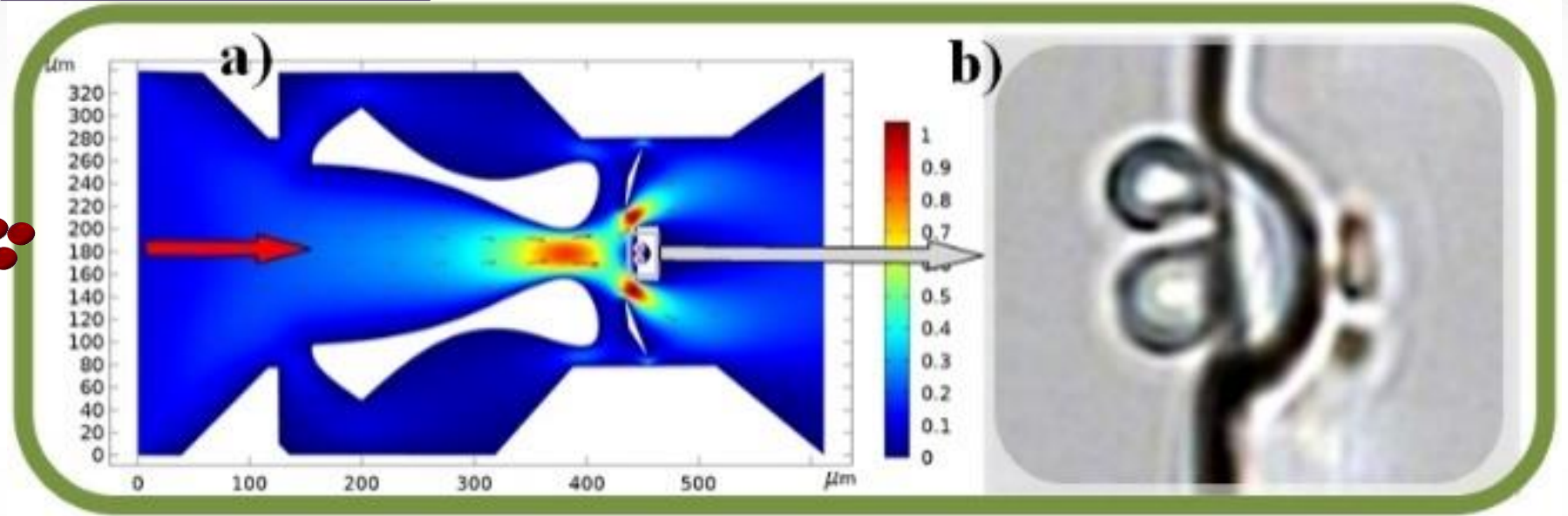
- Design and fabricate microfluidic chip suitable for LOT
- Run the RBC solution through the chip and trap at least one cell
- Mathematically restore the 3D surface of the trapped RBC (RBCs surface vary per RBC type)



## Raman spectrometry

- Design and fabricate microfluidic chip suitable for Raman spectrometry
- Run the RBC solution through the chip and trap at least one cell
- Obtain spectra of the chip + trapped RBC
- Clearly detect hemoglobin on the spectra

# Concept

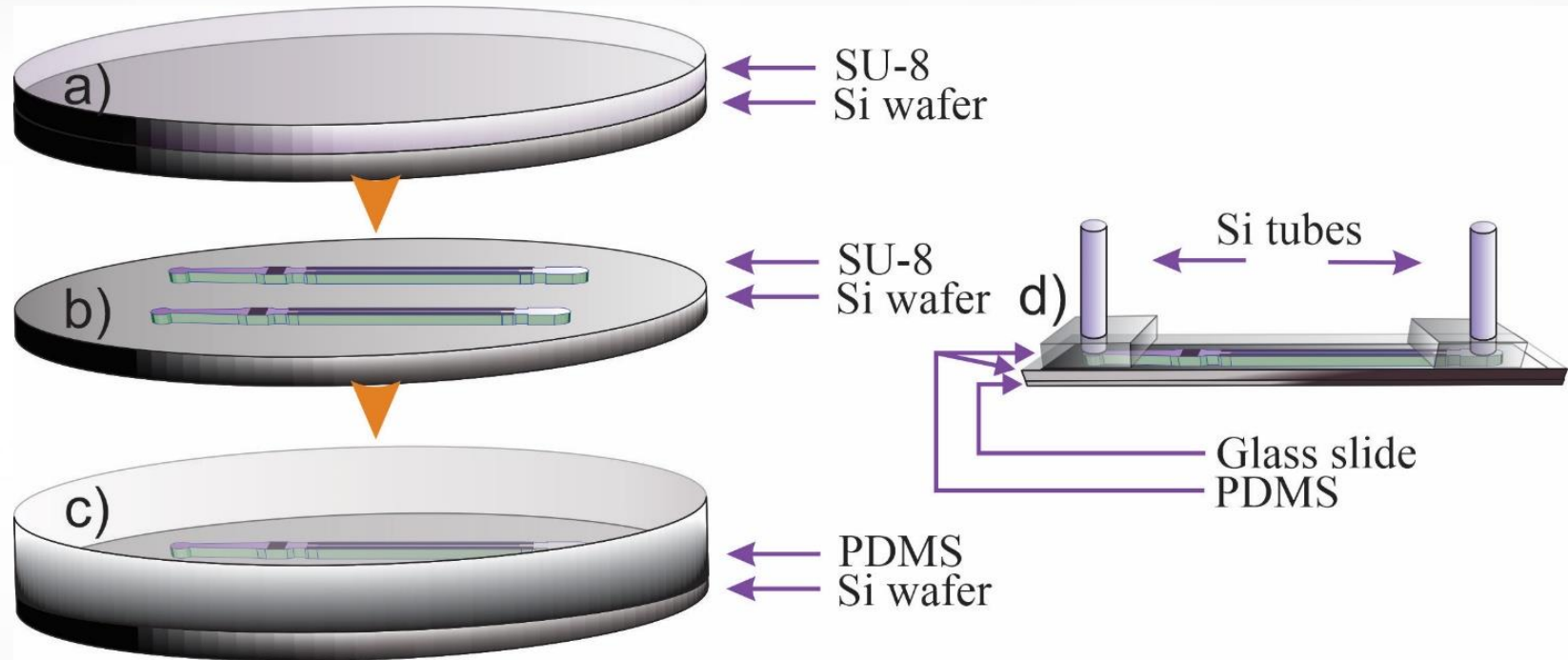


## Simulation and trapping.

a) COMSOL simulation of the velocity gradient distribution in the trapping chamber (inflow velocity = 0.1 m/s, no slip boundary condition), red arrow indicates the flow direction;

b) 2 RBCs trapped in the mid-section cavity of the chamber and ready for tests.

# Fabrication



**Figure: chip fabrication**

- a) photoresist spincoating; b) photolithography;  
c) PDMS mold; d) chip packaging

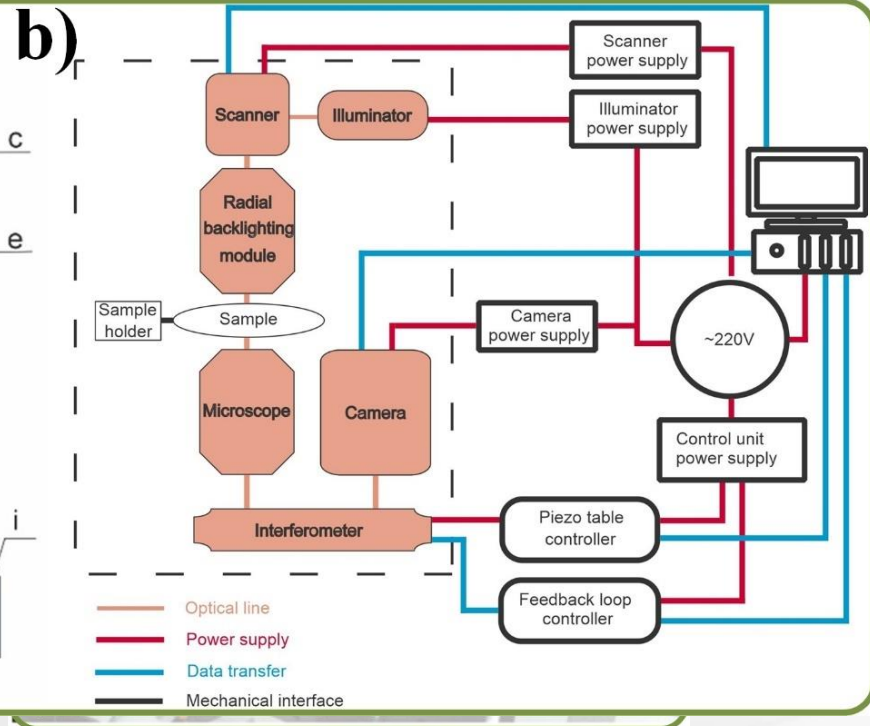


# Local optical tomography by differential projections



we have a  
an internal

Whole image from the DIC-projections (1) is proportional to the Hilbert transform of the original function of the object. Applying inverse Hilbert transform (2) to DIC-projections image the tomogram of an object is reconstructed [10.1134/S0030400X18120226].



$$b(x, z) = -2\pi H_x[f(x, z)]$$

$$f(x, z) = \frac{1}{2\pi} H_x[b(x, z)]$$

$$b(x, z) = \int_{-\frac{\pi}{2}}^{+\frac{\pi}{2}} g(x \cos \varphi + z \sin \varphi; \varphi) d\varphi$$

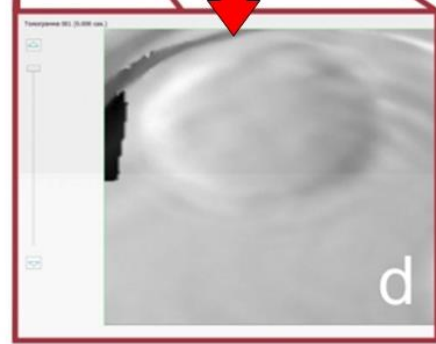
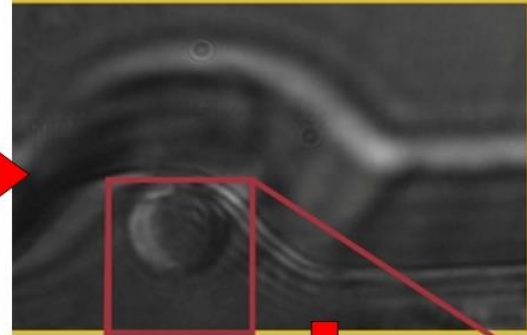
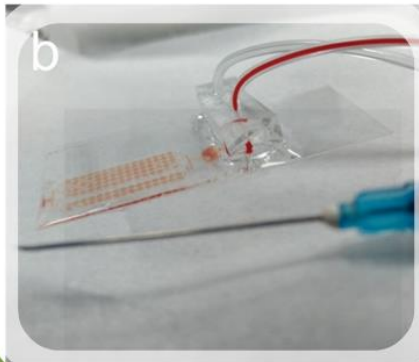
is total image from the DIC projections,  $f(x, z)$  is 2D image,

$\varphi$  is the angle between the perpendicular and the  $x$  axis on the interval  $[-\frac{\pi}{2}, +\frac{\pi}{2}]$

$H_x$  – Hilbert transform operator by  $x$ -axis and

$g$  is a partial derivative of the 1<sup>st</sup> order ( $r$ -coordinate) of central section

# LOT

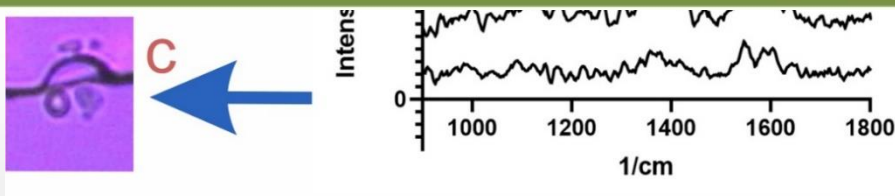
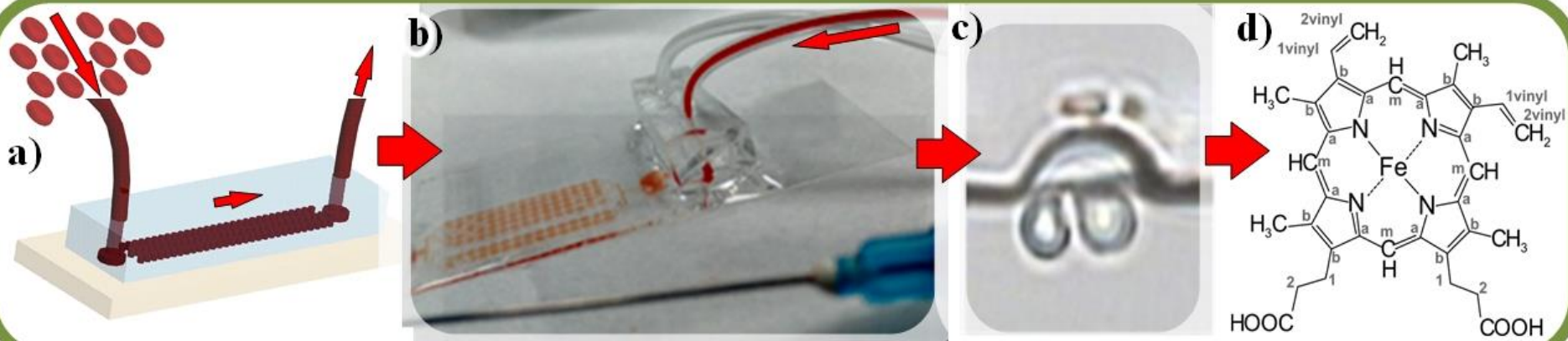
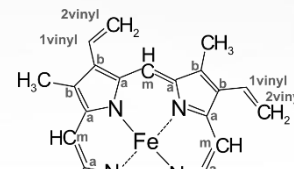


## *Typical fabricated microfluidic device*

*a) chip with one line used for blood solution being injected, due to the limitations of the Local Optical Tomograph the overall thickness is limited by 300  $\mu\text{m}$  and output flow should be open, having no tubes or secondary PDMS slabs on top of it; b) device thickness comparison to a 0.6 mm thick medical syringe injector needle; c) – an example of a PDMS first layer film used for LOT*

# Results of Raman spectrometry (combinatorial scattering)

Raman shift, $\text{cm}^{-1}$	Chemical bonds	The sensitivity of band
1640	$\text{C}_a\text{C}_m$ , $\text{C}_a\text{C}_m\text{H}$ , $\text{C}_a\text{C}_b$	Redox and spin state of Fe, presence of ligand
1588-1580	$\text{C}_a\text{C}_m$ , $\text{C}_a\text{C}_m\text{H}$	Spin state of heme Fe, diameter of porphyrin ring
1552	$\text{C}_a\text{C}_m$ , $\text{C}_a\text{C}_m\text{H}$	Spin state of heme Fe, diameter of porphyrin ring



# TOP. OPT. 1: OF MINIMIZATION. MANUAL CREATION/TESTING OF NEW GEOMETRIES

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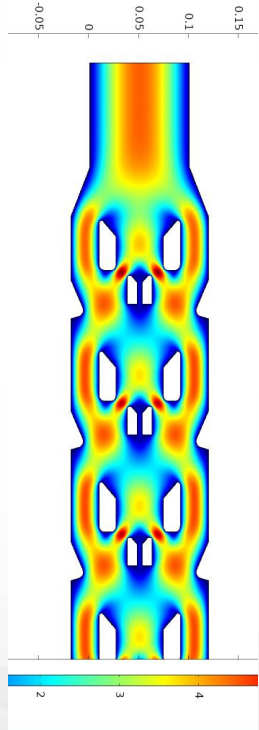


# Hydrodynamic

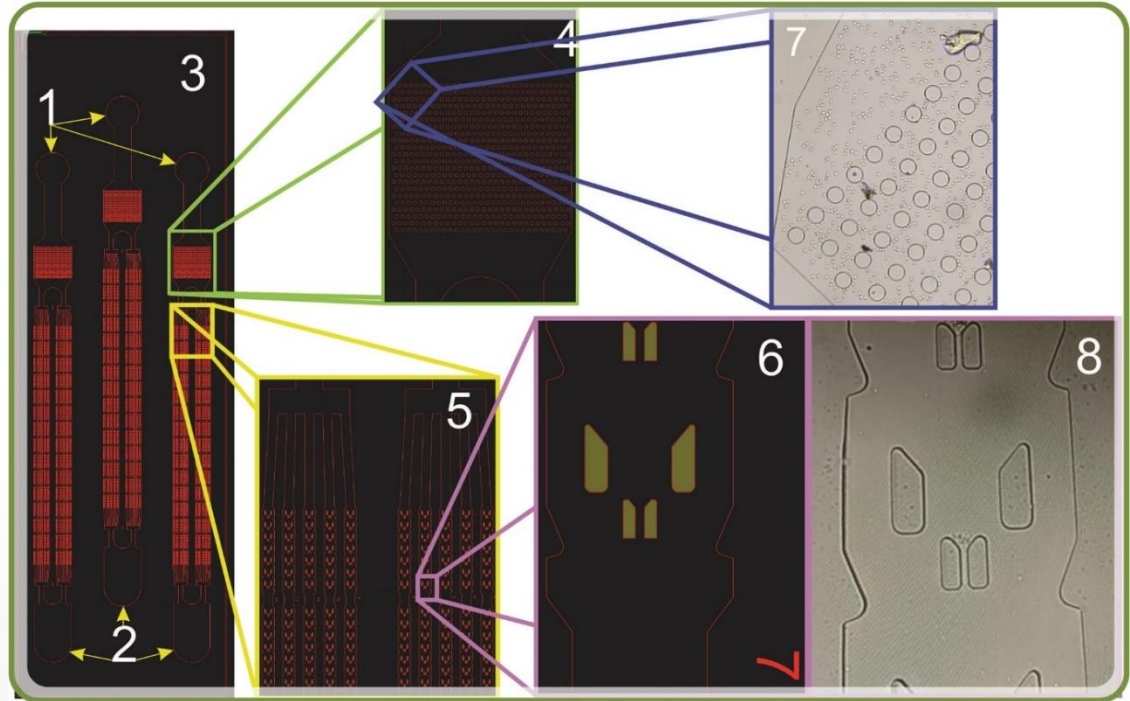
# RBC

# SiCMA

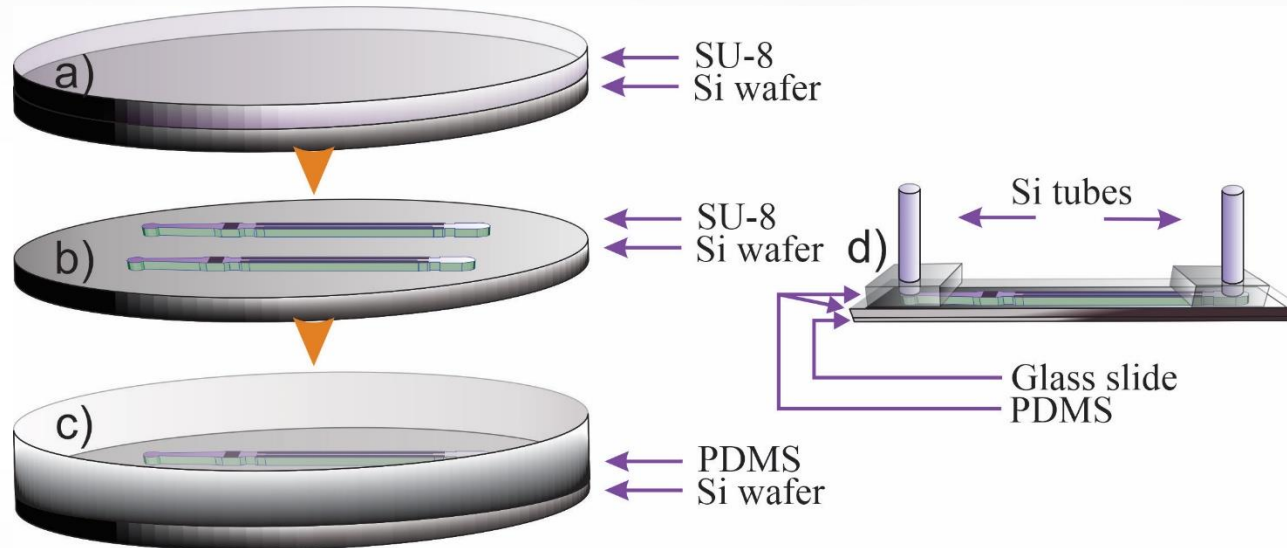
- **Design:** Glass (1.01; 0.17 mm) + PDMS (4.8; 0.13 mm) round chamber (1 mm), V-shaped trap, holding/capturing concavity  $d = 25/35$  micron (15 columns, 8 rows, 2 channels per chip == 240 chambers).



Typical geometry of the devices used in the experiments for establishing the optimal width of the trapping channels.



# Fabrication

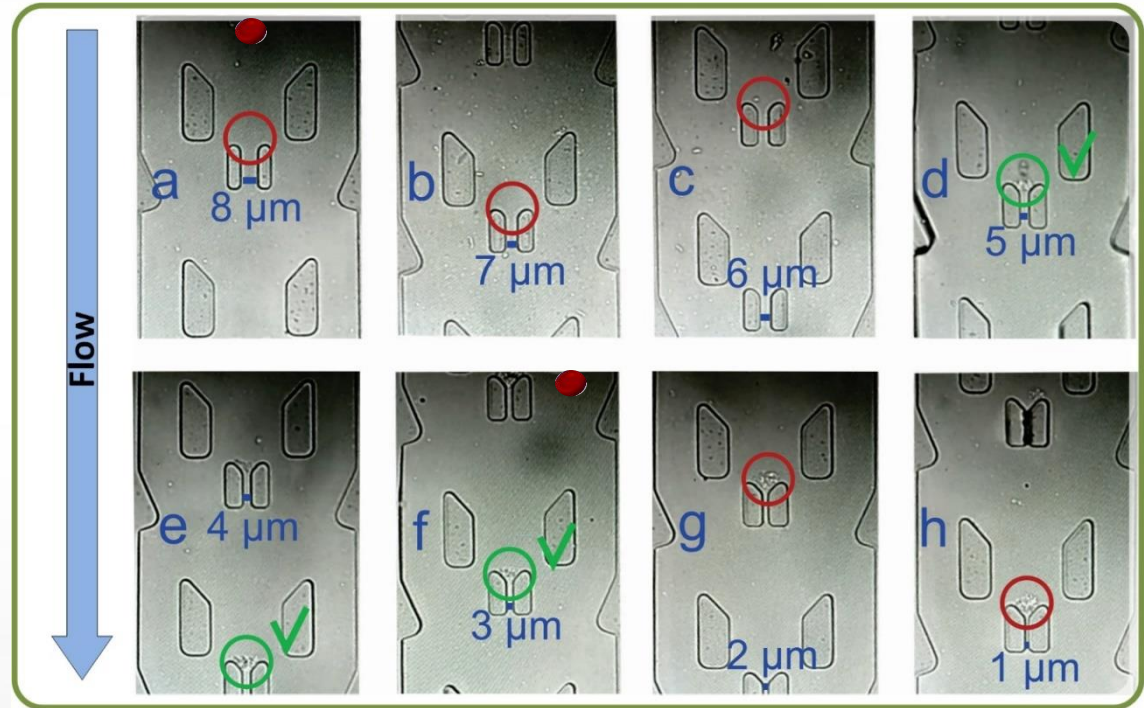


**Figure: chip fabrication**

a) photoresist spincoating; b) photolithography;  
c) PDMS mold; d) chip packaging

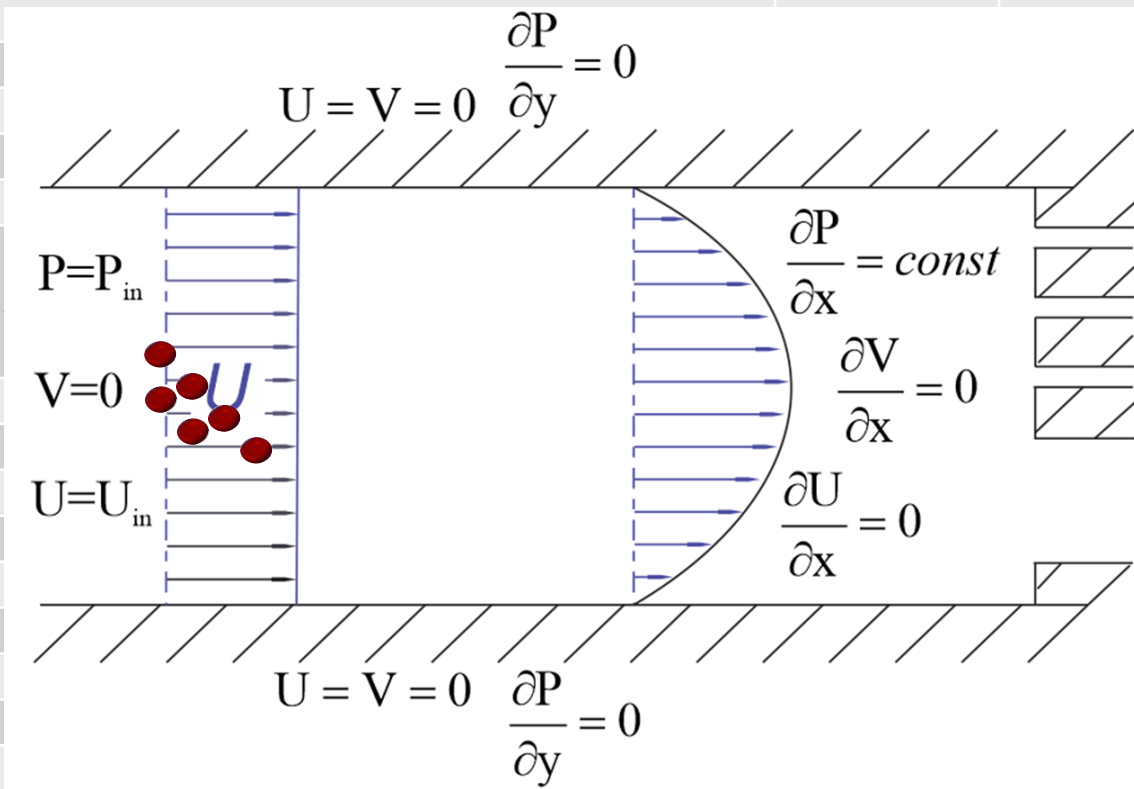
# Experimental microscopic images of establishing the optimal trapping width for trapping a target cell.

f, g, h – bovine erythrocytes, a, b, c, d, e – human RBCs. To find the optimal parameters of a trap different geometries were tested within one chip: width ranging from 1  $\mu\text{m}$  to 8  $\mu\text{m}$ . a) 8  $\mu\text{m}$ ; b) 7  $\mu\text{m}$ ; c) 6  $\mu\text{m}$ ; d) 5  $\mu\text{m}$ ; e) 4  $\mu\text{m}$ ; f) 3  $\mu\text{m}$ ; g) 2  $\mu\text{m}$ ; h) 1  $\mu\text{m}$ ; the diameter of the human RBC = 7-8  $\mu\text{m}$ <sup>462</sup>, the size of the bovine RBC = 5-6  $\mu\text{m}$ <sup>474</sup>



# Flow solution rheological parameters (blood density...)

f_large	intop1(u)	m <sup>2</sup> /s	Flow rate through the large outlet
f_1	intop2(u)	m <sup>2</sup> /s	Flow rate through the small outlet
f_2			Flow rate ratio
alpha			Friction force
target_f_large			Target flow rate 1
target_f_small			Target flow rate 2
volume_f			Volume fraction of the material
(f_1 - 1)^2			Objective function
obj			
f_3			
curl			
f_4			
f_5			
Re			Reynolds number
mu0			Dynamic viscosity
rho0			Density
D			Inlet width
Uin			Average inlet velocity
meshsz	0.0005[mm]	5E-7 m	Mesh size





Objective function

Flow rates, units =  $10^{-6} \text{ m}^2/\text{s}$

Broad channel  $F_0$

1-st channel

2-nd channel

3-rd channel

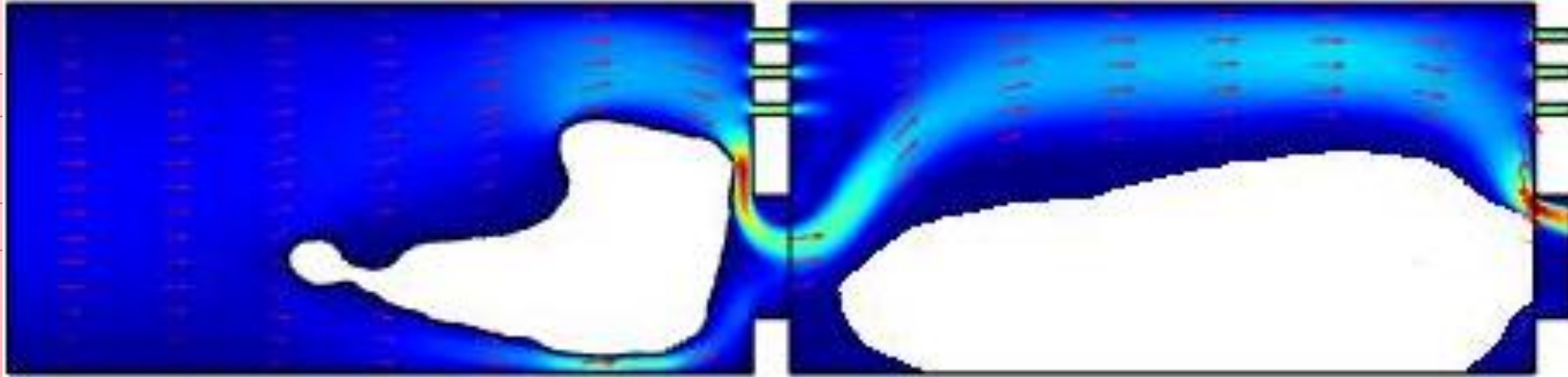
0 (optimization off)

95.8

1.37

1.38

1.39



$(F_1 / T_1 - 1)^2 + (F_2 / T_2 - 1)^2 + (F_3 / T_3 - 1)^2 + 4(F_0 / T_0 - 1)^2$

58.9

13.7

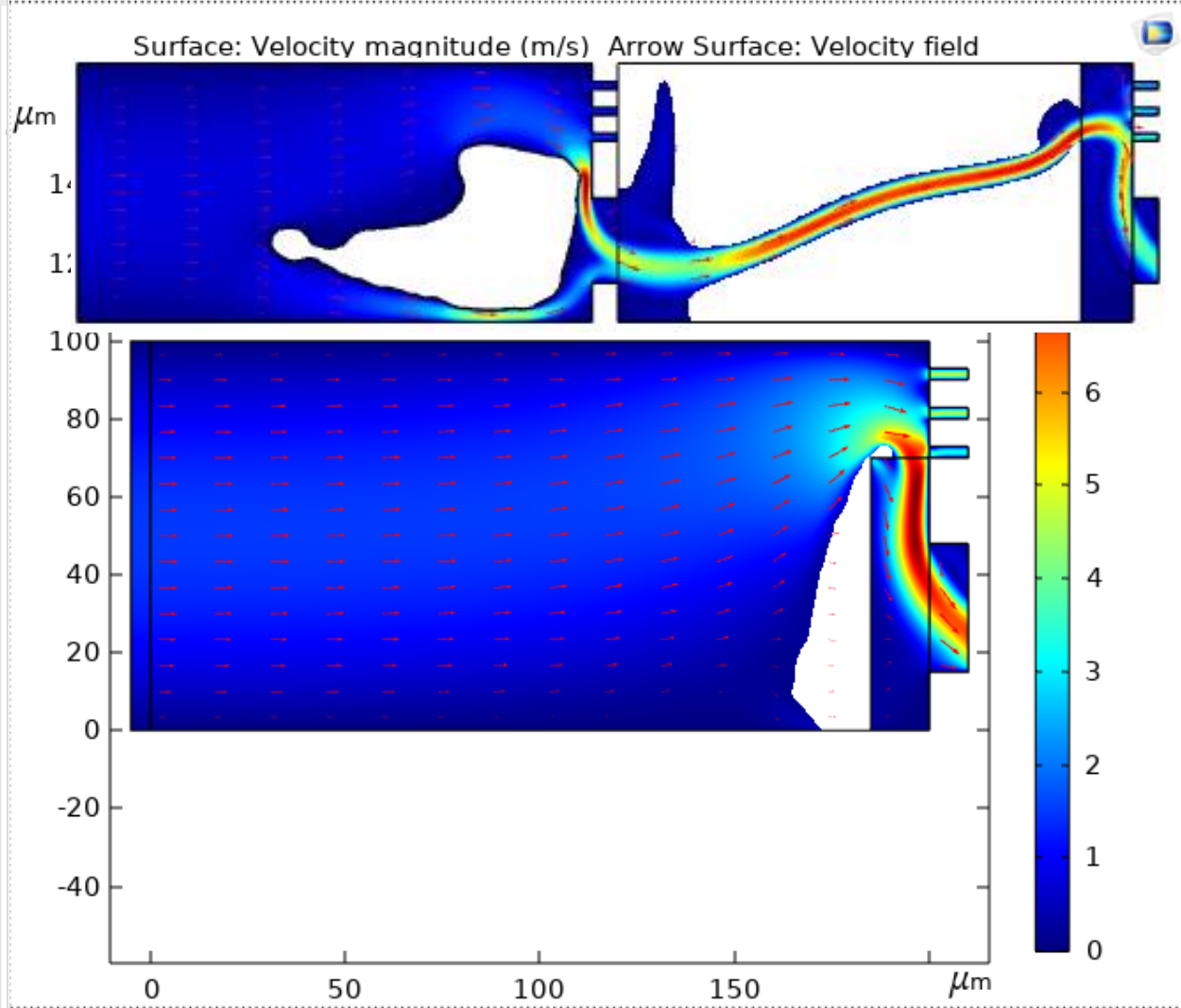
13.7

13.7

$T_1 = T_2 = T_3 = 40.0$

$T_0 = 100.0$

$$= \left( \frac{\int_{\text{narrow\_channel}_1} u \, dl}{\text{target}_{f_{\text{narrow}}}} - 1 \right)^2 + \left( \frac{\int_{\text{narrow\_channel}_2} u \, dl}{\text{target}_{f_{\text{narrow}}}} - 1 \right)^2 + \left( \frac{\int_{\text{narrow\_channel}_3} u \, dl}{\text{target}_{f_{\text{narrow}}}} - 1 \right)^2 + 4 * \left( \frac{\int_{\text{broad}} dl}{\text{target}_{f_{\text{broad}}}} - 1 \right)^2 \quad (3.1)$$



# Constraints

$$CRV = \iint abs \frac{u^2 * \frac{\partial v}{\partial x} - v^2 \frac{\partial u}{\partial y} + uv * \left( \frac{\partial v}{\partial y} - \frac{\partial u}{\partial x} \right)}{(u^2 + v^2)^{3/2}} dx dy$$

OF

$$= \left( \frac{\int_{narrow\_channel\_1} u \, dl}{target_{f_{narrow}}} - 1 \right)^2 + \left( \frac{\int_{narrow\_channel\_2} u \, dl}{target_{f_{narrow}}} - 1 \right)^2$$

$$+ \left( \frac{\int_{narrow\_channel\_3} u \, dl}{target_{f_{narrow}}} - 1 \right)^2 + \left( \frac{\int_{narrow\_channel\_4} u \, dl}{target_{f_{narrow}}} - 1 \right)^2$$

$$+ \left( \frac{\int_{narrow\_channel\_5} u \, dl}{target_{f_{narrow}}} - 1 \right)^2 + k * \left( \frac{f_{broad} dl}{target_{f_{broad}}} - 1 \right)^2$$

OF

$$\begin{aligned} &= \left( \frac{\int_{\text{narrow\_channel}_1} u \, dl}{\text{target}_{f_{\text{narrow}}}} - 1 \right)^2 + \left( \frac{\int_{\text{narrow\_channel}_2} u \, dl}{\text{target}_{f_{\text{narrow}}}} - 1 \right)^2 \\ &+ \left( \frac{\int_{\text{narrow\_channel}_3} u \, dl}{\text{target}_{f_{\text{narrow}}}} - 1 \right)^2 + \left( \frac{\int_{\text{narrow\_channel}_4} u \, dl}{\text{target}_{f_{\text{narrow}}}} - 1 \right)^2 \\ &+ \left( \frac{\int_{\text{narrow\_channel}_5} u \, dl}{\text{target}_{f_{\text{narrow}}}} - 1 \right)^2 + k * \left( \frac{f_{\text{broad}} dl}{\text{target}_{f_{\text{broad}}}} - 1 \right)^2 \\ &+ \left( \frac{\max_{\Omega} \left( \frac{\partial u}{\partial y} \right) + \max_{\Omega} \left( \frac{\partial v}{\partial x} \right)}{\text{target}_{\text{CRL}}} - 1 \right)^2 \\ &+ \left( \frac{\iint_{\Omega} \text{abs} \left[ \frac{u^2 * \frac{\partial v}{\partial x} - v^2 \frac{\partial u}{\partial y} + uv * \left( \frac{\partial v}{\partial y} - \frac{\partial u}{\partial x} \right)}{(u^2 + v^2)^{3/2}} \right] dx dy}{\text{target}_{\text{CRV}}} - 1 \right)^2 \end{aligned}$$

# Constraints

$$CRL = \maxop1(uy) + \maxop1(vx)$$

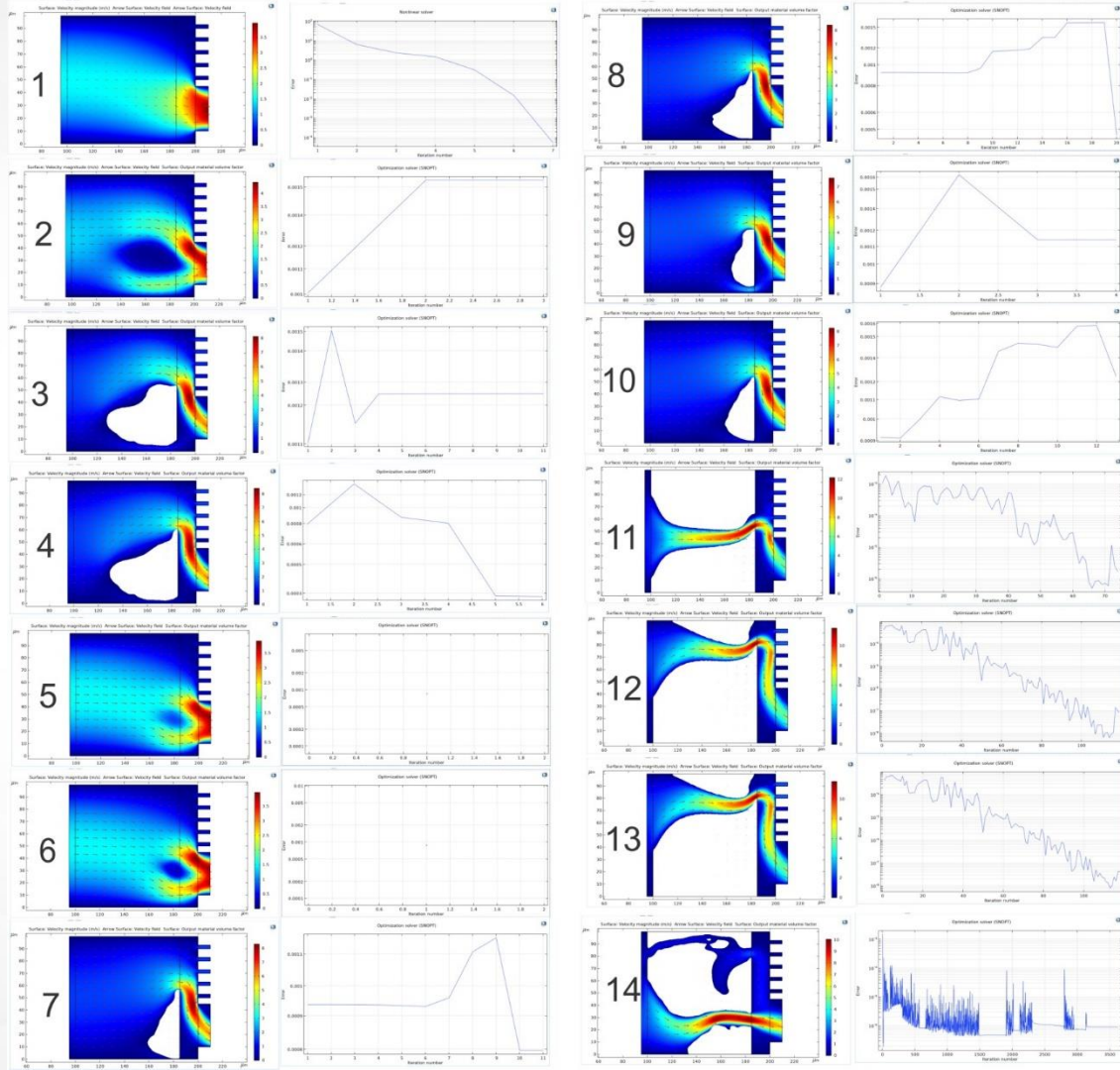
$$\max_{\Omega} \left( \frac{\partial u}{\partial y} \right) + \max_{\Omega} \left( \frac{\partial v}{\partial x} \right)$$

$$CRV = \intop1(abs(u^2 * vx - v^2 * uy + u*v*(vy - ux))/(u^2 + v^2)^{1.5})$$

$$\iint_{\Omega} abs \left[ \frac{u^2 * \frac{\partial v}{\partial x} - v^2 \frac{\partial u}{\partial y} + uv * \left( \frac{\partial v}{\partial y} - \frac{\partial u}{\partial x} \right)}{(u^2 + v^2)^{3/2}} \right] dx dy$$



# OF topology optimization



# Pareto front parametrical plot

coefficients for the parts of the OF			Target values				Probed value after the simulation					simulation NN
F_Broad	curl	curvature	target_f_small	target_curl	target_curvature	target_f_broad	F_1	F_5	curl	curvature	F_broad	
4.0	1.0	8.0	4E-5[m^2/s]	2.00E+07	2E8[1/m]	1E-4[m^2/s]	5.96E-07	5.46E-07	3.23E+06	1.00E+08	9.71E-05	1
3.0	2.0	5.0	4E-5[m^2/s]	2.00E+07	2E8[1/m]	1E-4[m^2/s]	1.06E-06	8.20E-07	8.51E+06	2.06E+08	9.50E-05	2
2.0	4.0	2.0	4E-5[m^2/s]	2.00E+07	2E8[1/m]	1E-4[m^2/s]	2.98E-06	1.36E-06	1.11E+07	1.50E+08	8.72E-05	3
8.0	8.0	0.5	4E-5[m^2/s]	2.00E+07	2E8[1/m]	1E-4[m^2/s]	3.65E-06	1.09E-06	1.12E+07	2.25E+08	8.53E-05	4
2.0	2.0	15.0	4E-5[m^2/s]	2.00E+07	2E8[1/m]	1E-4[m^2/s]	8.03E-07	7.25E-07	7.90E+06	1.50E+08	9.60E-05	5
0.5	20.0	40.0	4E-5[m^2/s]	2.00E+07	2E8[1/m]	1E-4[m^2/s]	8.72E-07	7.79E-07	8.54E+06	9.43E+07	9.57E-05	6
0.5	2.0	4.0	4E-5[m^2/s]	2.00E+07	2E8[1/m]	1E-4[m^2/s]	3.44E-06	1.39E-06	1.18E+07	1.82E+08	8.54E-05	7
0.2	4.0	6.0	4E-5[m^2/s]	2.00E+07	2E8[1/m]	1E-4[m^2/s]	3.45E-06	1.52E-06	1.21E+07	1.95E+08	8.51E-05	8
0.1	10.0	8.0	4E-5[m^2/s]	2.00E+07	2E8[1/m]	1E-4[m^2/s]	2.68E-06	1.47E-06	1.10E+07	9.90E+07	8.80E-05	9
0.6	0.7	0.8	4E-5[m^2/s]	2.00E+07	2E8[1/m]	1E-4[m^2/s]	3.34E-06	1.53E-06	1.20E+07	1.72E+08	8.55E-05	10
1.0	0.0	0.0	4E-5[m^2/s]	2.00E+07	2E8[1/m]	1E-4[m^2/s]	4.22E-06	3.94E-06	1.84E+07	2.70E+08	7.85E-05	11
1.0	0.0	0.0	4E-5[m^2/s]	2.00E+07	2E8[1/m]	1E-4[m^2/s]	5.58E-06	5.79E-07	1.43E+07	3.54E+07	8.42E-05	12
1.0	0.0	0.0	4E-5[m^2/s]	2.00E+07	2E8[1/m]	1E-4[m^2/s]	5.58E-06	5.79E-07	1.43E+07	3.54E+07	8.42E-05	13
1.0	0.0	0.0	4E-6[m^2/s]	2.00E+07	2E8[1/m]	1E-5[m^2/s]	4.22E-06	3.92E-06	1.84E+07	1.42E+08	7.85E-05	14
1.0	0.0	0.0	4E-7[m^2/s]	2.00E+07	2E8[1/m]	1E-3[m^2/s]	4.00E-07	3.58E-08	3.66E+06	1.10E+08	9.88E-05	15
1.0	1.0	1.0	4E-6[m^2/s]	2.00E+07	2E8[1/m]	1E-4[m^2/s]	3.50E-06	1.04E-06	1.09E+07	2.00E+08	8.59E-05	16
1.0	20.0	20.0	4E-4[m^2/s]	2.00E+07	2E8[1/m]	1E-6[m^2/s]	4.21E-06	3.97E-06	1.85E+07	2.14E+08	7.85E-05	17



**TOP. OPT. 2: AUTO  
CREATION/TESTING  
OF NEW GEOMETRIES  
(10, 000 DATASETS).**

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# Constraints & variables

## Constraints:

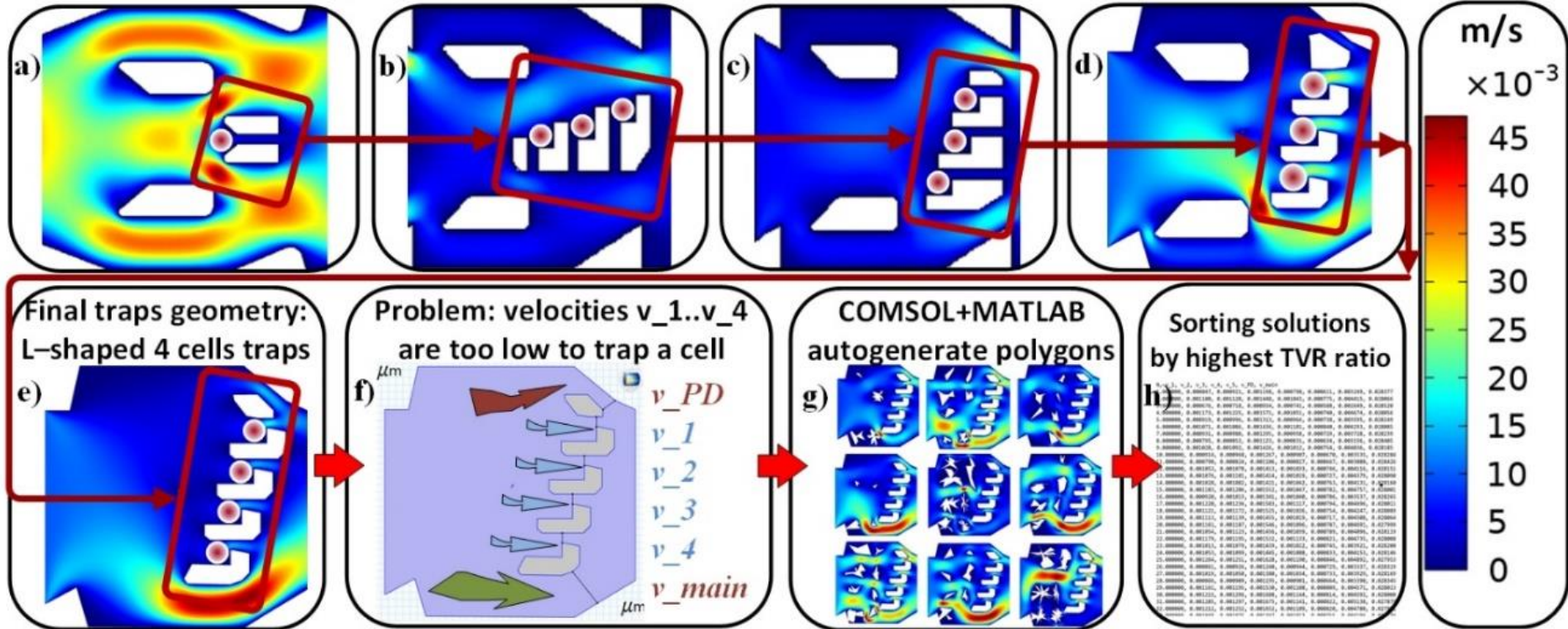
- flow curvature, flow width, flow rates.
- flow curvature (no flow direction rapid changes);
- flow width within the channel/chamber (the limit is - not narrower than  $\frac{1}{4}$  or  $\frac{1}{3}$  of “broad channel” width);
- flow rates (each “narrow outlet” has  $\frac{1}{3}$  of a “broad outlet” flow rate).

## Parameters that may vary within a range:

- width of the whole chamber (50 ... 250 microns);
- length of the whole chamber (10 ... 200 microns) excluding outlets length;
- number of narrow outlet channels (3 ... 15);
- width of the broad outlet channel (50 ... 100 microns);
- length of the narrow and broad outlets (2 ... 50 microns).

# Topology Optimization 2

The design stages of cell traps and architecture of the evolutionary algorithm



N, v\_1, v\_2, v\_3, v\_4, v\_5, v\_PD, v\_main

1.000000	0.000847	0.000921	0.001198	0.000798	0.000611	0.003249	0.028377
2.000000	0.001100	0.001120	0.001448	0.001043	0.000775	0.004415	0.028066
3.000000	0.000676	0.000718	0.000954	0.000741	0.000588	0.002649	0.028520
4.000000	0.001173	0.001225	0.001571	0.001051	0.000740	0.004674	0.028056
5.000000	0.000919	0.000996	0.001313	0.000964	0.000738	0.003595	0.028249
6.000000	0.001071	0.001086	0.001436	0.001101	0.000840	0.004293	0.028085
7.000000	0.000931	0.000980	0.001295	0.000958	0.000729	0.003728	0.028239
8.000000	0.000795	0.000852	0.001123	0.000831	0.000634	0.003156	0.028405
9.000000	0.001028	0.001092	0.001426	0.001012	0.000754	0.004036	0.028185
10.000000	0.000914	0.000968	0.001267	0.000907	0.000670	0.003535	0.028284
11.000000	0.000790	0.000824	0.001106	0.000827	0.000667	0.003080	0.028426
12.000000	0.001053	0.001078	0.001413	0.001033	0.000746	0.004116	0.028151
13.000000	0.001076	0.001101	0.001414	0.001019	0.000727	0.004379	0.028090
14.000000	0.001028	0.001082	0.001421	0.001042	0.000763	0.004131	0.028160
15.000000	0.001183	0.001206	0.001552	0.001047	0.000782	0.004757	0.028001
16.000000	0.000920	0.001013	0.001341	0.001040	0.000786	0.003537	0.028265
17.000000	0.001220	0.001236	0.001583	0.001117	0.000794	0.004696	0.028011
18.000000	0.001121	0.001172	0.001525	0.001026	0.000754	0.004247	0.028089
19.000000	0.001113	0.001139	0.001455	0.001019	0.000717	0.004508	0.028064
20.000000	0.001161	0.001187	0.001546	0.001096	0.000787	0.004691	0.027999
21.000000	0.001054	0.001123	0.001456	0.001039	0.000789	0.004096	0.028119
22.000000	0.001179	0.001195	0.001532	0.001133	0.000821	0.004735	0.028008
23.000000	0.001013	0.001079	0.001439	0.001022	0.000745	0.003921	0.028200
24.000000	0.001053	0.001099	0.001445	0.001088	0.000833	0.004153	0.028146
25.000000	0.001264	0.001251	0.001628	0.001190	0.000846	0.004892	0.027953
26.000000	0.000861	0.000926	0.001240	0.000944	0.000729	0.003337	0.028319
27.000000	0.001019	0.001050	0.001380	0.001034	0.000733	0.003929	0.028169
28.000000	0.000866	0.000909	0.001195	0.000901	0.000664	0.003390	0.028345
29.000000	0.001161	0.001195	0.001530	0.001108	0.000809	0.004571	0.028013
30.000000	0.001215	0.001296	0.001688	0.001144	0.000814	0.004592	0.028000
31.000000	0.001285	0.001297	0.001675	0.001141	0.000822	0.005138	0.027875
32.000000	0.001212	0.001252	0.001652	0.001109	0.000820	0.004780	0.027971
33.000000	0.001048	0.001075	0.001307	0.001013	0.000755	0.004195	0.028135

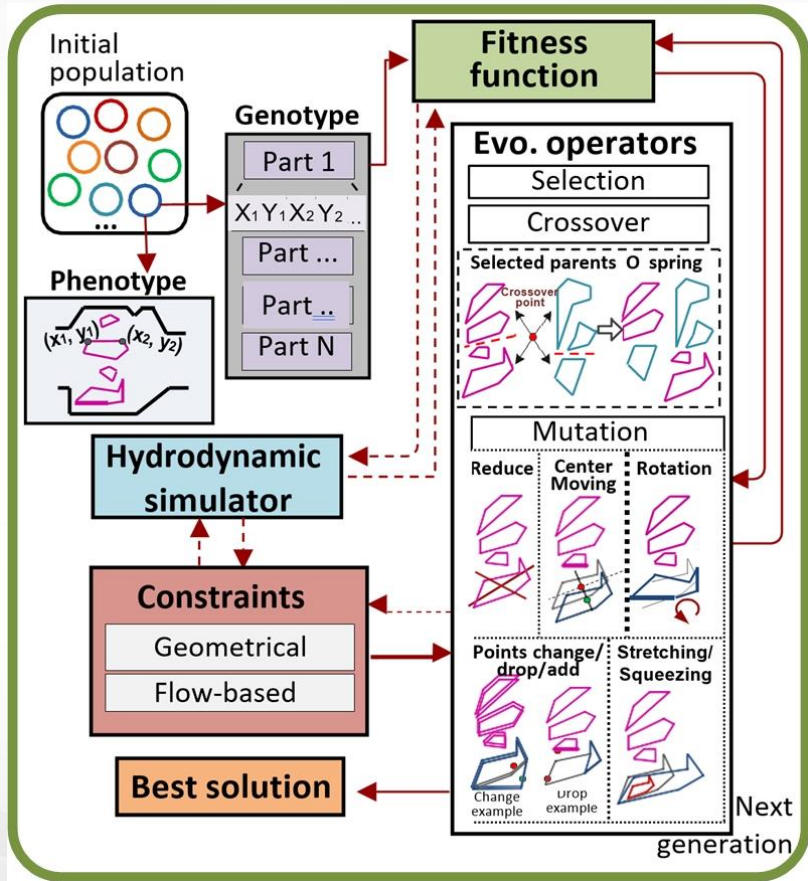


# TOP. OPT. 3: EA, AUTO CREATION AND IMPROVEMENT OF 300 NEW GEOMETRIES

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# Generative design of microfluidic structures using evolutionary approach 1



The scheme of evolutionary approach for the design of microfluidic geometry and an example of problem-specific evolutionary operators.

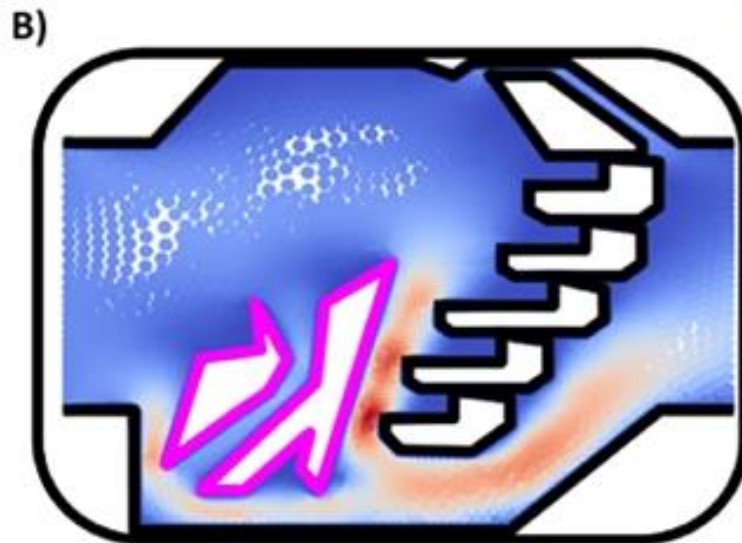
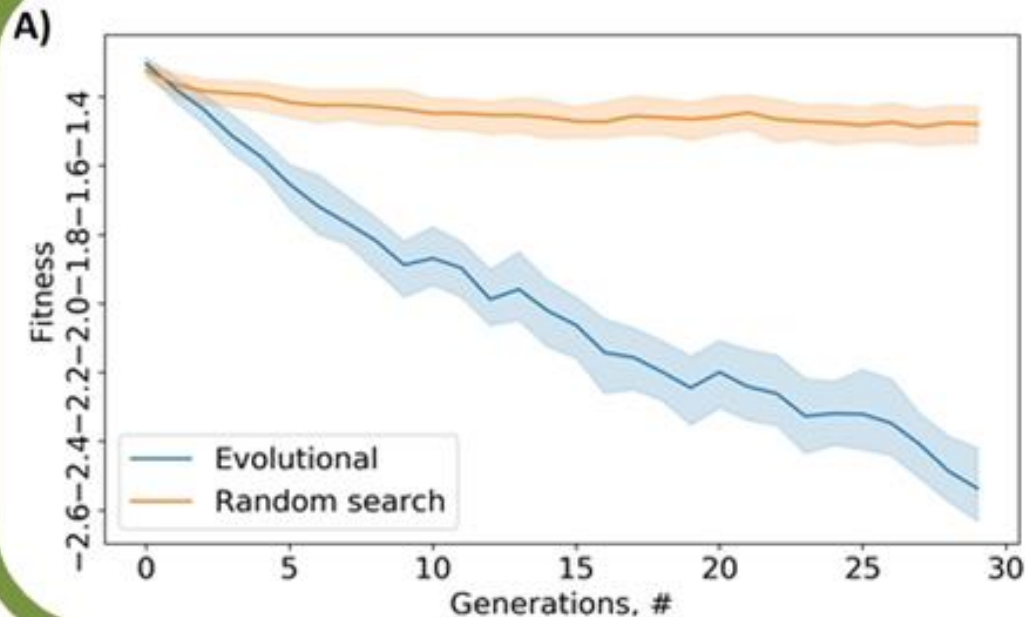
$$X_{opt} = \arg \min_X F(X),$$

$$F(X) = G(F(Y(X)) | G(X)), G(X) = (g_1(X), \dots, g_M(X))$$

$$X = (P_1, P_2, \dots, P_N), P = (Pt_1, Pt_2, \dots, Pt_{ki}), Pt_j = (x_j, y_j)$$

# Topology Optimization 3

target=0.258867 curv=1.657296 curl=24217.406313 width\_ratio=0.33712



pop\_num

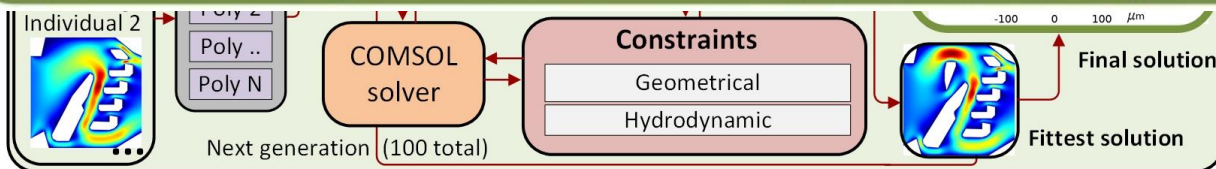
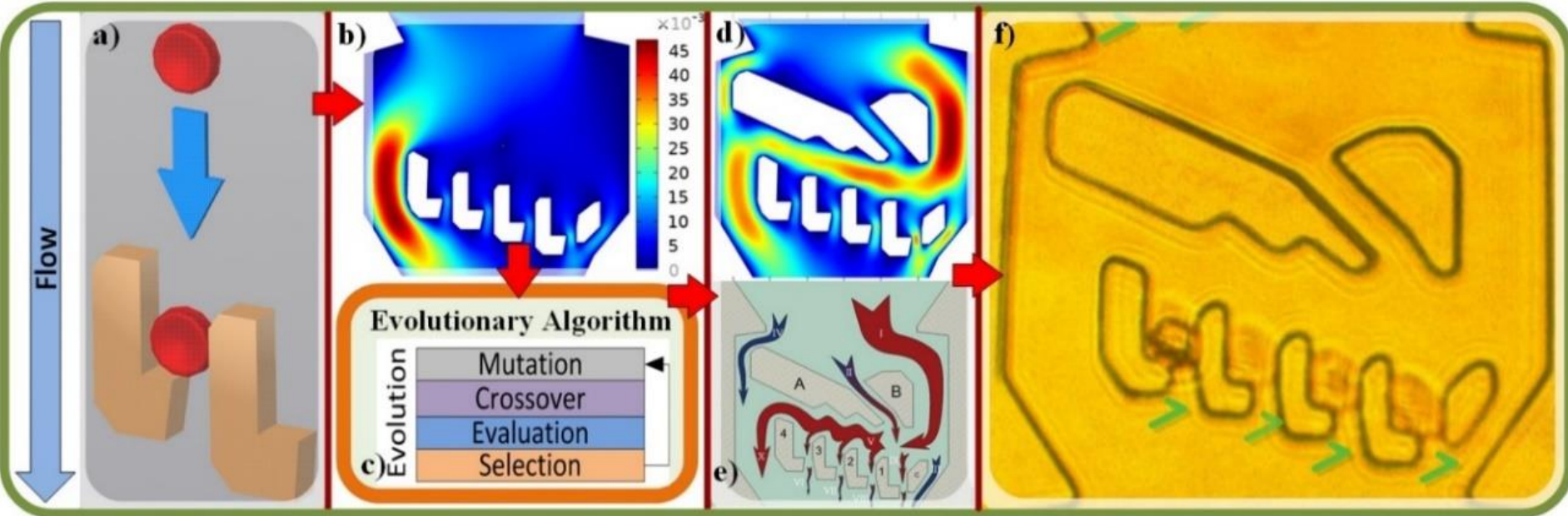
**TOP. OPT. 4: EA, AUTO  
CREATION SELF-  
IMPROVEMENT OF 30,  
000 NEW GEOMETRIES**

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# Design stages of cell traps and the architecture of the evolutionary algorithm

The design stages of cell traps and architecture of the evolutionary algorithm





# Constraints

$$CRL = \maxop1(uy) + \maxop1(vx)$$

$$\max_{\Omega} \left( \frac{\partial u}{\partial y} \right) + \max_{\Omega} \left( \frac{\partial v}{\partial x} \right)$$

$$CRV = \text{intop1}(\text{abs}(u^2 * vx - v^2 * uy + u*v*(vy - ux))/(u^2 + v^2)^{1.5})$$

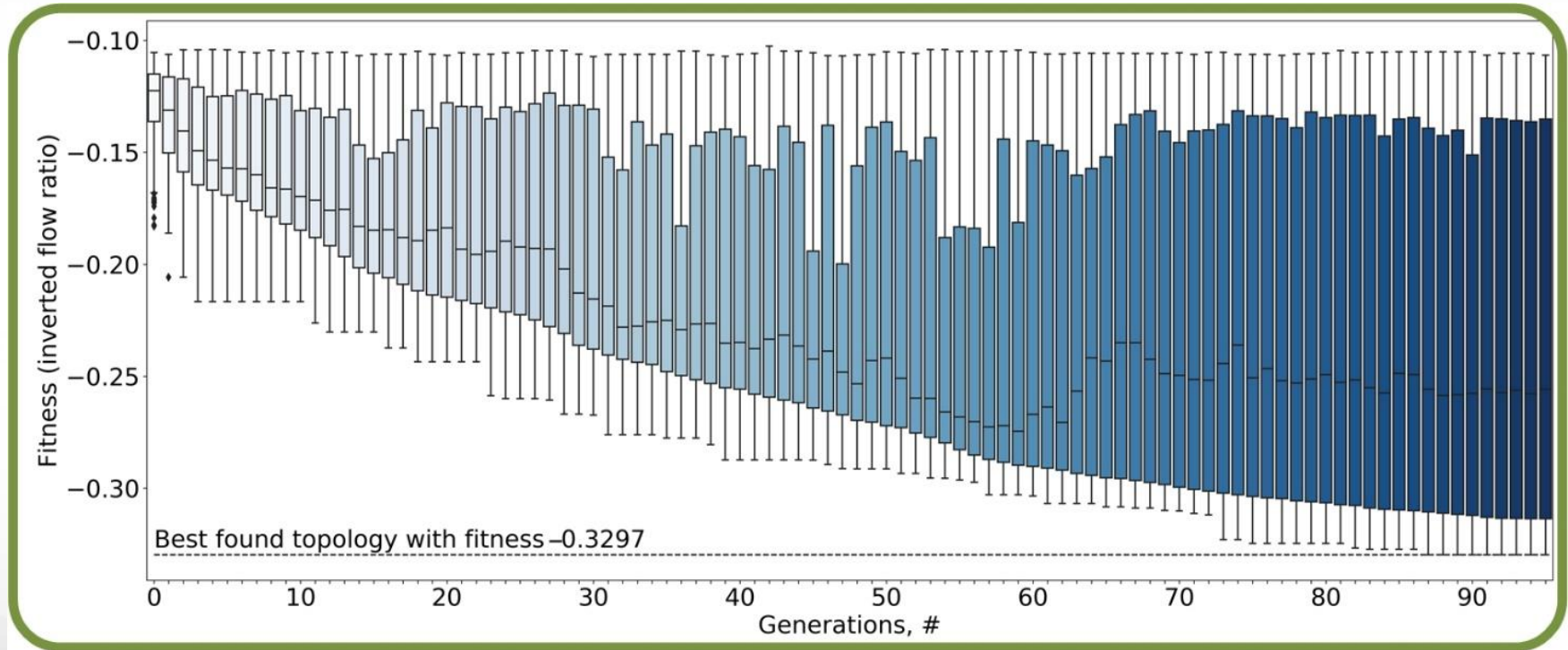
$$\iint_{\Omega} \text{abs} \left[ \frac{u^2 * \frac{\partial v}{\partial x} - v^2 \frac{\partial u}{\partial y} + uv * \left( \frac{\partial v}{\partial y} - \frac{\partial u}{\partial x} \right)}{(u^2 + v^2)^{3/2}} \right] dx dy$$

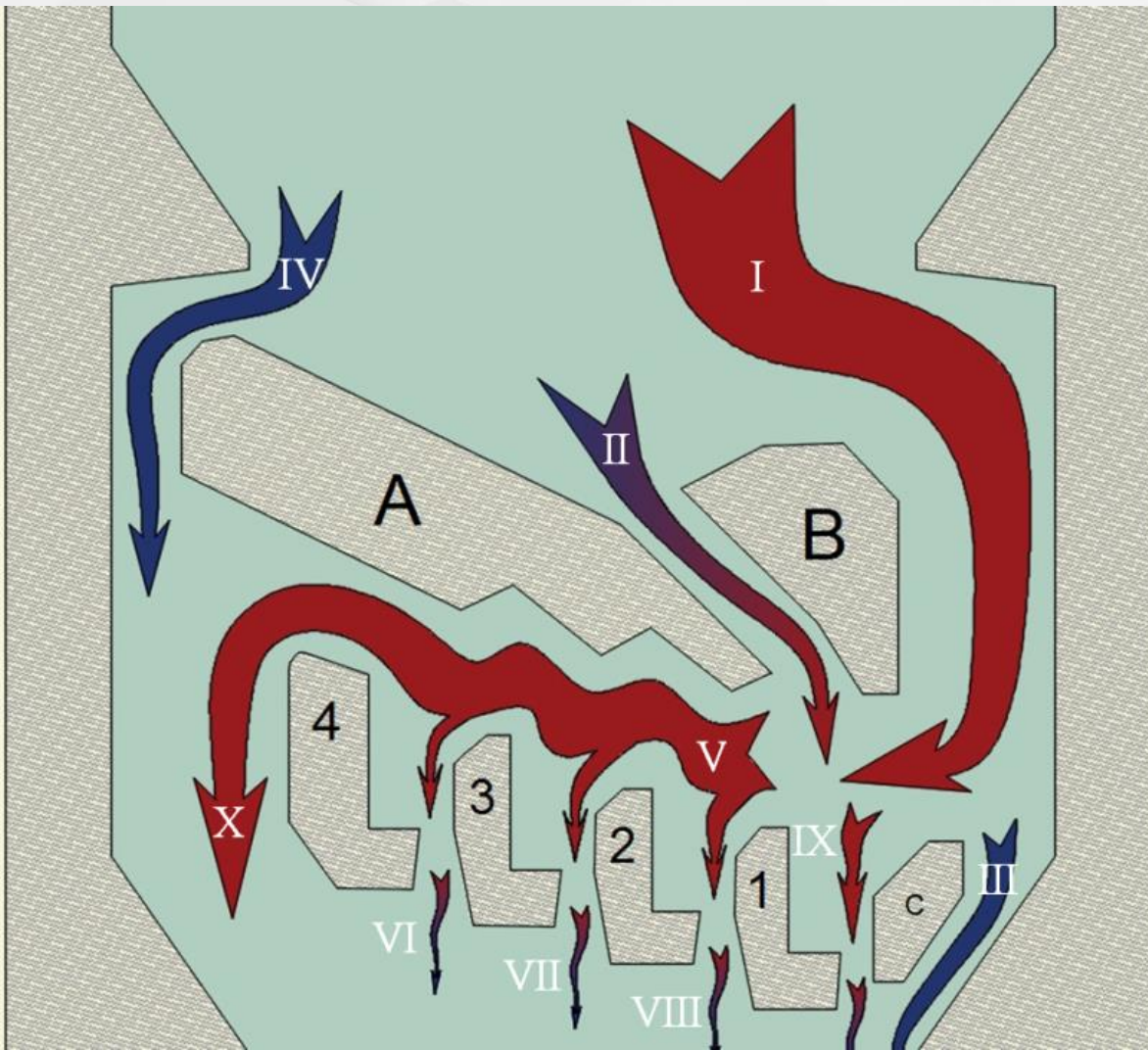
$$TVR = \frac{\sum_{k=1}^4 v_k}{v_{PD} + v_{main}}$$



# Convergence graph

*The convergence of the fitness function values (the function that is used to estimate how close a given design solution is to the specified aim) during the evolutionary optimization of cell traps for the 100 generations (iterations of evolution).*





*Final optimized geometry of the microfluidic trap for single cell*

# Experimental results of the system following the evolutionary algorithm design to trap RBCs.

Parameter	Units	Initial	Target values	Optimized	Gain, %
vl_1	m/s	0.012038	determined	0.02308	<b>92</b>
vl_2	m/s	0.0094433	by	0.01579	<b>67</b>
vl_3	m/s	0.0094776		0.012701	<b>34</b>
vl_4	m/s	0.0095439		0.010092	<b>6</b>
vl_PD	m/s	0.0059983		TVR ratio	0.012438
vl_main	m/s	0.027247		0.019577	<b>-28</b>
CVR	l/m	70,769,000	$< 7 \times 10^{+07}$	17,113,000	<b>-76</b>
CRL	l/s	12717	$< 30,000$	20615	<b>62</b>
TVR (target)	-	<b>1.22</b>	<b><math>1.22 &lt; \text{TVR} &lt; 2</math></b>	<b>1.93</b>	<b>58</b>

— flow velocity, um/s

— cell trapping probability, %

— flow velocity, um/s

— cell trapping probability, %

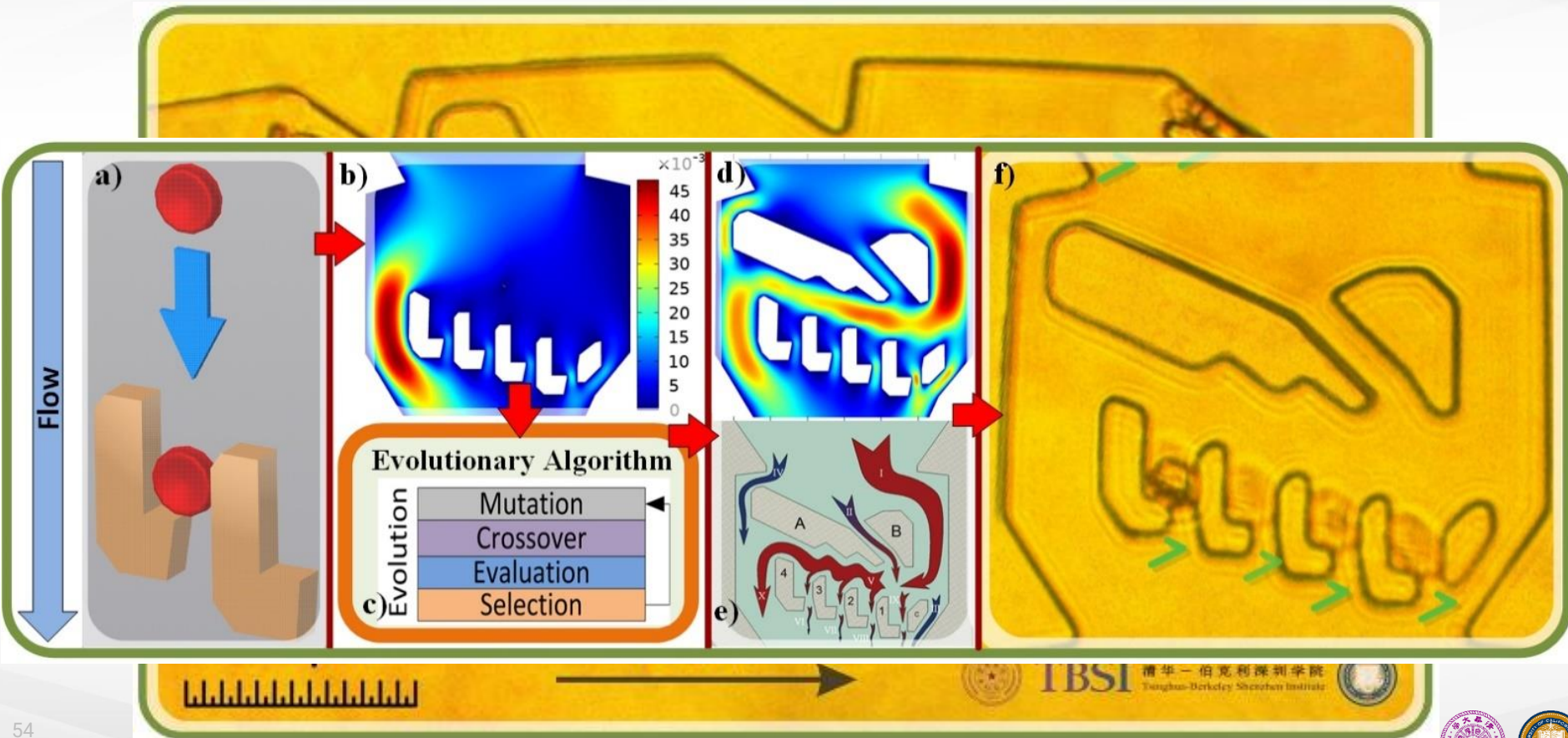


# CONCLUSION AND CLOSING REMARKS

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# uFD design overview

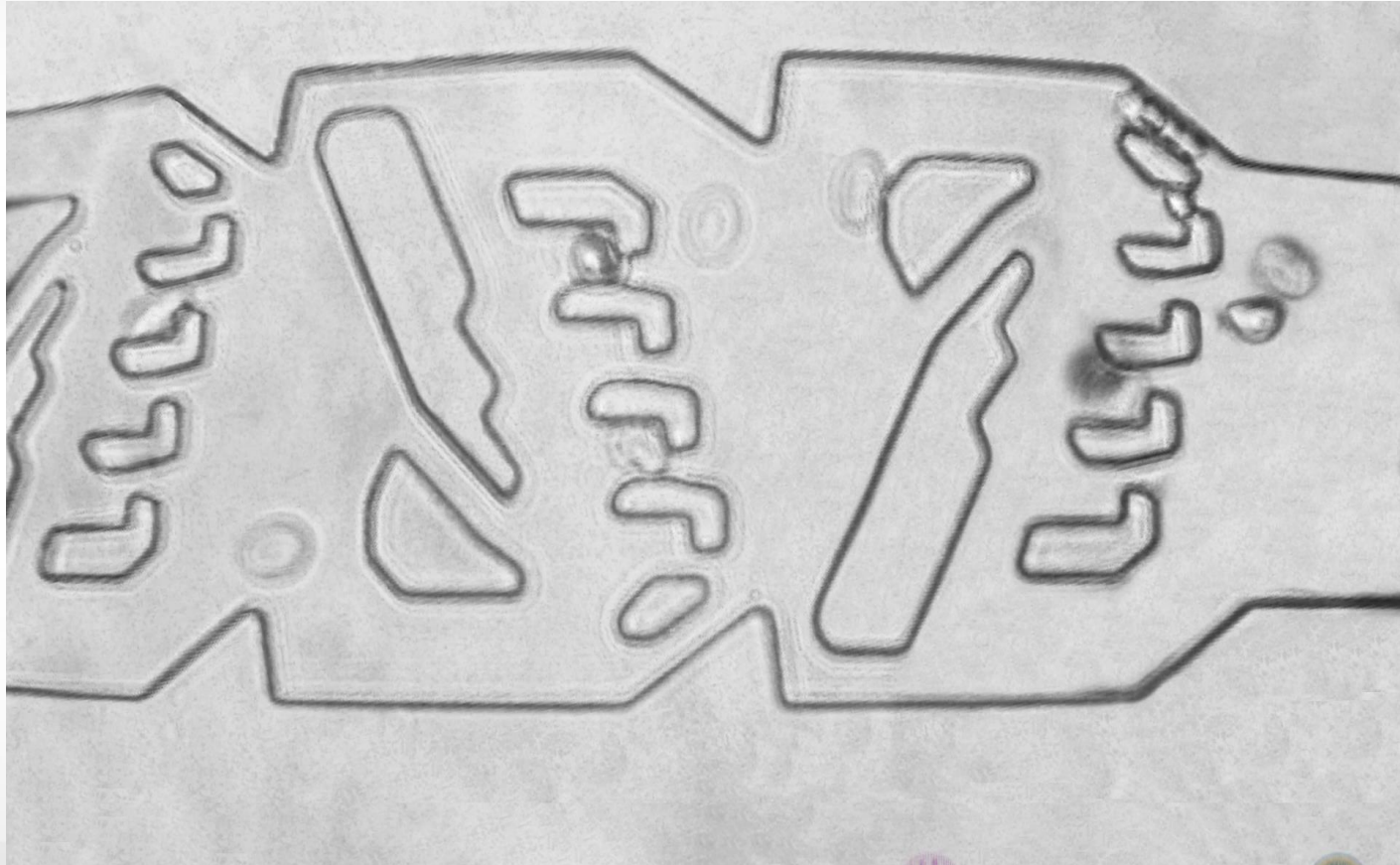


# Discussion

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# uFD successfully trapping cells





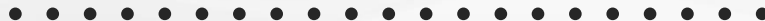
# References

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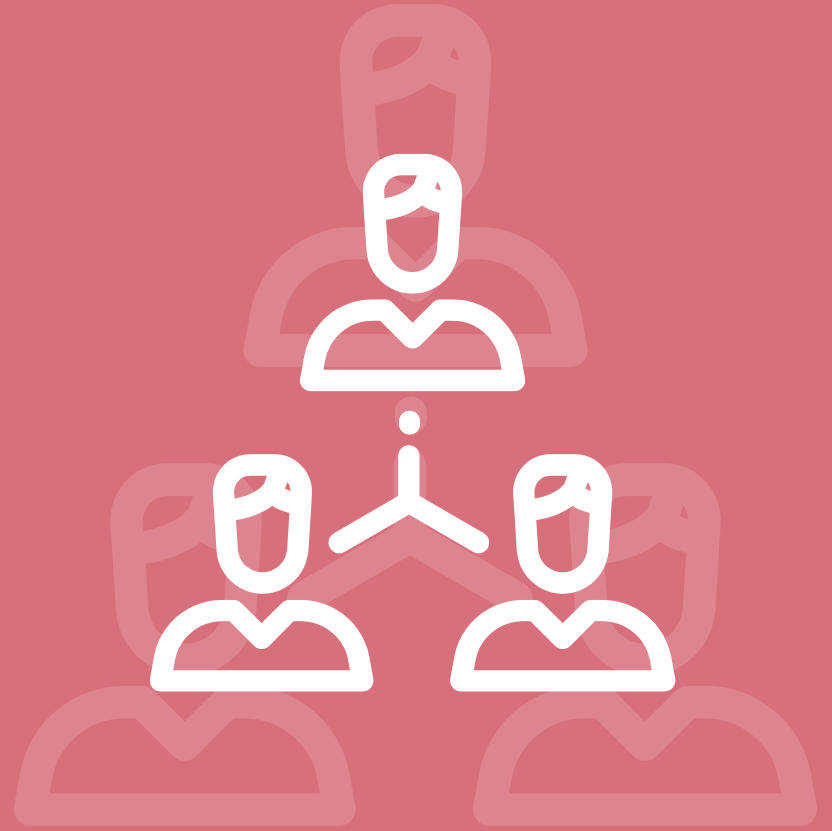
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# Collaborations

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- *Suchalko Oleg Ivanovich*, Master student;

**ВНИИОФИ, УИ**

- *Visl*
- *L*
- *E*

**НИИО  
Acaden**

- *Me*
- *Ko*

**ИТМО**

- *Ka*
- *Ni*
- *Hv*

**中国科**

- *Sh*
- *Fe*

**深圳北**

- *M*
- *T*

**Stanfor**

- *Sayk*
- *Kapitu*

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biology” of Russian

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DLOGY



ITMO UNIVERSITY

ogical Sciences;



# Future work

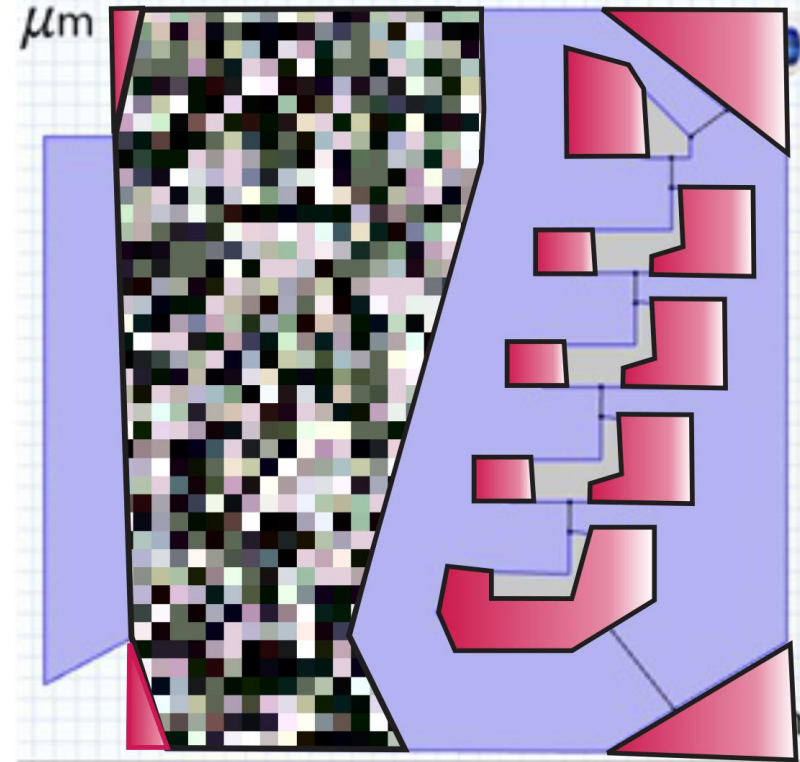
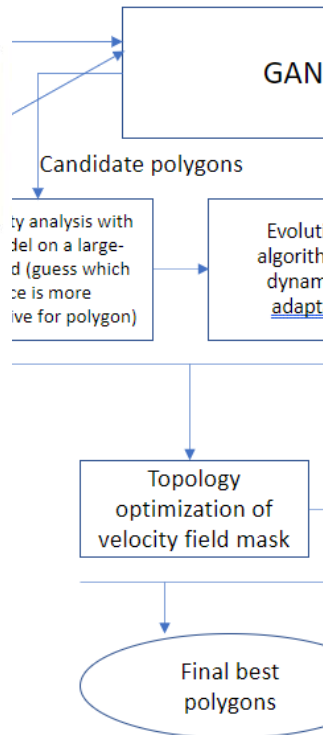
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# Future development of EA based on Generative Adversarial Networks



nal optimization  
st solutions



**Благодарю за  
внимание!**

**懇請各位老師批評指正**

